

Linuron
Analysis of Risks
to
Endangered and Threatened Salmon and Steelhead

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Summary

Linuron is a substituted urea herbicide registered for use on asparagus, carrots, celery, corn (field and sweet), cottonseed (use voluntarily removed by registrant), parsley, parsnips, potatoes, sorghum, soybeans, and winter wheat. Linuron may be applied preplant, preemergence, postemergence, or post transplant using ground equipment. The registered modes of application are band treatment, directed spray, or broadcast spray. The most current EPA records show 9 active end-use product registrations and 5 technical grades of linuron. End-use products include wettable powder (50% a.i.), flowable concentrate (40.6% a.i.), water dispersible granules (50% a.i.), and liquid suspensions.

Linuron functions as an herbicide through the inhibition of photosynthesis. It is approved for a wide range of noxious weeds, but local conditions and weed resistance have limited its use to specific states and, often, specific portions of states. These combined factors greatly reduce actual linuron use from levels provided for in EPA registration documents and registrant labels. Toxicity to fish is considered slight to moderate, however linuron is quite toxic to aquatic invertebrates. The invertebrate toxicity is probably the most significant potential adverse effect from this pesticide since invertebrates serve as a primary food source for young and juvenile salmon and steelhead. A potential concern, addressed in the attached specific analysis of the ESUs, is the concentration of linuron use in a few counties associated with the spawning and rearing territories of salmon and steelhead. A general presumption is that young organisms are more sensitive to any adverse influence than mature organisms. This concern is reflected in the conclusions regarding linuron use in California and the Pacific Northwest.

The main use of linuron is to control germinating and newly emerging grasses and broad leaf weeds in soybeans. Generally, it is applied to newly emerging crops as an over the top spray. In asparagus, it is applied between cutting of newly emerging spears for weed control during harvest.

¹ Comment: Data and the analysis based upon it reflects information available at the time this report was completed. Additional data, which may be submitted or change in status after the submission date are not included in the authors evaluations, presentations, or comments.

Scope - Although this analysis is specific to listed western salmon and steelhead and the watersheds in which they occur, it is acknowledged that linuron is registered for uses that may occur outside this geographic scope and that additional analyses may be required to address other T&E species in the Pacific states as well as across the United States. I understand that any subsequent analyses, requests for consultation, and resulting Biological Opinions may necessitate that Biological Opinions relative to this request be revisited, and could be modified. Much of the quantitative information presented and used was derived from the Registration Eligibility Decision (RED; Attachment 1) and the Ecological Risk Assessment (ERA, Attachment 2) developed by the Ecological Fate and Effects Division (EFED) for the RED.

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1. Background

Under section 7 of the Endangered Species Act, the Office of Pesticide Programs (OPP) of the U. S. Environmental Protection Agency (EPA) is required to consult on actions that may affect Federally listed endangered or threatened species or that may adversely modify designated critical habitat. Situations where a pesticide may affect a fish, such as any of the salmonid species listed by the National Marine Fisheries Service (NMFS), include either direct or indirect

effects on the fish. Direct effects result from exposure to a pesticide at levels that may cause harm.

Acute Toxicity - Relevant acute data are derived from standardized toxicity tests with lethality as the primary endpoint. These tests are conducted with what is generally accepted as the most sensitive life stage of fish, i.e., very young fish from 0.5-5 grams in weight, and with species that are usually among the most sensitive. These tests for pesticide registration include analysis of observable sublethal effects as well. The intent of acute tests is to statistically derive a median effect level; typically the effect is lethality in fish (LC50) or immobility in aquatic invertebrates (EC50). Typically, a standard fish acute test will include concentrations that cause no mortality, and often no observable sublethal effects, as well as concentrations that would cause 100% mortality. By looking at the effects at various test concentrations, a dose-response curve can be derived, and one can statistically predict the effects likely to occur at various pesticide concentrations; a well done test can even be extrapolated, with caution, to concentrations below those tested (or above the test concentrations if the highest concentration did not produce 100% mortality).

OPP typically uses qualitative descriptors to describe different levels of acute toxicity, the most likely kind of effect of modern pesticides (Table 1). These are widely used for comparative purposes, but must be associated with exposure before any conclusions can be drawn with respect to risk. Pesticides that are considered highly toxic or very highly toxic are required to have a label statement indicating that level of toxicity. The FIFRA regulations [40CFR158.490(a)] do not require calculating a specific LC50 or EC50 for pesticides that are practically non-toxic; the LC50 or EC50 would simply be expressed as >100 ppm. When no lethal or sublethal effects are observed at 100 ppm, OPP considers the pesticide will have “no effect” on the species.

Table 1. Qualitative descriptors for categories of fish and aquatic invertebrate toxicity (from Zucker, 1985)

LC50 or EC50	Category description
< 0.1 ppm	Very highly toxic
0.1- 1 ppm	Highly toxic
>1 < 10 ppm	Moderately toxic
> 10 < 100 ppm	Slightly toxic
> 100 ppm	Practically non-toxic

Comparative toxicology has demonstrated that various species of scaled fish generally have equivalent sensitivity, within an order of magnitude, to other species of scaled fish tested under the same conditions. Exceptions are known to occur for only an occasional pesticide, as based on the several dozen fish species that have been frequently tested. Sappington et al.

(2001), Beyers et al. (1994) and Dwyer et al. (1999), among others, have shown that endangered and threatened fish tested to date are similarly sensitive, on an acute basis, to a variety of pesticides and other chemicals as are their non-endangered counterparts.

Chronic Toxicity - OPP evaluates the potential chronic effects of a pesticide on the basis of several types of tests. These tests are often required for registration, but not always. If a pesticide has essentially no acute toxicity at relevant concentrations, or if it degrades very rapidly in water, or if the nature of the use is such that the pesticide will not reach water, then chronic fish tests may not be required [40CFR158.490]. Chronic fish tests primarily evaluate the potential for reproductive effects and effects on the offspring. Other observed sublethal effects are also required to be reported. An abbreviated chronic test, the fish early-life stage test, is usually the first chronic test conducted and will indicate the likelihood of reproductive or chronic effects at relevant concentrations. If such effects are found, then a full fish life-cycle test will be conducted. If the nature of the chemical is such that reproductive effects are expected, the abbreviated test may be skipped in favor of the full life-cycle test. These chronic tests are designed to determine a “no observable effect level” (NOEL) and a “lowest observable effect level” (LOEL). A chronic risk requires not only chronic toxicity, but also chronic exposure, which can result from a chemical being persistent and resident in an environment (e.g., a pond) for a chronic period of time or from repeated applications that transport into any environment such that exposure would be considered “chronic”.

As with comparative toxicology efforts relative to sensitivity for acute effects, EPA, in conjunction with the U. S. Geological Survey, has a current effort to assess the comparative toxicology for chronic effects also. Preliminary information indicates, as with the acute data, that endangered and threatened fish are again of similar sensitivity to similar non-endangered species.

Metabolites and Degradates - Information must be reported to OPP regarding any pesticide metabolites or degradates that may pose a toxicological risk or that may persist in the environment [40CFR159.179]. Toxicity and/or persistence test data on such compounds may be required if, during the risk assessment, the nature of the metabolite or degradate and the amount that may occur in the environment raises a concern. If actual data or structure-activity analyses are not available, the requirement for testing is based upon best professional judgement.

Inert Ingredients - OPP does take into account the potential effects of what used to be termed “inert” ingredients, but which are beginning to be referred to as “other ingredients”. OPP has classified these ingredients into several categories. A few of these, such as nonylphenol, can no longer be used without including them on the label with a specific statement indicating the potential toxicity. Based upon our internal databases, I can find no product in which nonylphenol is now an ingredient. Many others, including such ingredients as clay, soybean oil, many polymers, and chlorophyll, have been evaluated through structure-activity analysis or data and determined to be of minimal or no toxicity. There exist also two additional lists, one for inerts with potential toxicity which are considered a testing priority, and one for inerts unlikely to be toxic, but which cannot yet be said to have negligible toxicity. Any new inert ingredients

are required to undergo testing unless it can be demonstrated that testing is unnecessary.

The inert efforts in OPP are oriented only towards toxicity at the present time, rather than risk. It should be noted, however, that very many of the inerts are in exceedingly small amounts in pesticide products. While some surfactants, solvents, and other ingredients may be present in fairly large amounts in various products, many are present only to a minor extent. These include such things as coloring agents, fragrances, and even the printers ink on water soluble bags of pesticides. Some of these could have moderate toxicity, yet still be of no consequence because of the negligible amounts present in a product. If a product contains inert ingredients in sufficient quantity to be of concern, relative to the toxicity of the active ingredient, OPP attempts to evaluate the potential effects of these inerts through data or structure-activity analysis, where necessary.

For a number of major pesticide products, testing has been conducted on the formulated end-use products that are used by the applicator. The results of fish toxicity tests with formulated products can be compared with the results of tests on the same species with the active ingredient only. A comparison of the results should indicate comparable sensitivity, relative to the percentage of active ingredient in the technical versus formulated product, if there is no extra activity due to the combination of inert ingredients. I note that the “comparable” sensitivity must take into account the natural variation in toxicity tests, which is up to 2-fold for the same species in the same laboratory under the same conditions, and which can be somewhat higher between different laboratories, especially when different stocks of test fish are used.

The comparison of formulated product and technical ingredient test results may not provide specific information on the individual inert ingredients, but rather is like a “black box” which sums up the effects of all ingredients. I consider this approach to be more appropriate than testing each individual inert and active ingredient because it incorporates any additivity, antagonism, and synergism effects that may occur and which might not be correctly evaluated from tests on the individual ingredients. I do note, however, that we do not have aquatic data on most formulated products, although we often have testing on one or perhaps two formulations of an active ingredient.

Risk - An analysis of toxicity, whether acute or chronic, lethal or sublethal, must be combined with an analysis of how much will be in the water, to determine risks to fish. Risk is a combination of exposure and toxicity. Even a very highly toxic chemical will not pose a risk if there is no exposure, or very minimal exposure relative to the toxicity. OPP uses a variety of chemical fate and transport data to develop “estimated environmental concentrations” (EECs) from a suite of established models. The development of aquatic EECs is a tiered process.

The first tier screening model for EECs is with the GENEEC program, developed within OPP, which uses a generic site (in Yazoo, MS) to stand for any site in the U. S. The site choice was intended to yield a maximum exposure, or “worst-case,” scenario applicable nationwide, particularly with respect to runoff. The model is based on a 10 hectare watershed that surrounds a one hectare pond, two meters deep. It is assumed that all of the 10 hectare area is treated with

the pesticide and that any runoff would drain into the pond. The model also incorporates spray drift, the amount of which is dependent primarily upon the droplet size of the spray. OPP assumes that if this model indicates no concerns when compared with the appropriate toxicity data, then further analysis is not necessary as there would be no effect on the species.

It should be noted that prior to the development of the GENEEC model in 1995, a much more crude approach was used to determining EECs. Older reviews and Reregistration Eligibility Decisions (REDs) may use this approach, but it was excessively conservative and does not provide a sound basis for modern risk assessments. For the purposes of endangered species consultations, we will attempt to revise this old approach with the GENEEC model, where the old screening level raised risk concerns.

When there is a concern with the comparison of toxicity with the EECs identified in GENEEC model, a more sophisticated PRZM-EXAMS model is run to refine the EECs if a suitable scenario has been developed and validated. The PRZM-EXAMS model was developed with widespread collaboration and review by chemical fate and transport experts, soil scientists, and agronomists throughout academia, government, and industry, where it is in common use. As with the GENEEC model, the basic model remains as a 10 hectare field surrounding and draining into a 1 hectare pond. Crop scenarios have been developed by OPP for specific sites, and the model uses site-specific data on soils, climate (especially precipitation), and the crop or site. Typically, site-scenarios are developed to provide for a worst-case analysis for a particular crop in a particular geographic region. The development of site scenarios is very time consuming; scenarios have not yet been developed for a number of crops and locations. OPP attempts to match the crop(s) under consideration with the most appropriate scenario. For some of the older OPP analyses, a very limited number of scenarios were available. As more scenarios become available and are geographically appropriate to selected T&E species, older models used in previous analyses may be updated.

One area of significant weakness in modeling EECs relates to residential uses, especially by homeowners, but also to an extent by commercial applicators. There are no usage data in OPP that relate to pesticide use by homeowners on a geographic scale that would be appropriate for an assessment of risks to listed species. For example, we may know the maximum application rate for a lawn pesticide, but we do not know the size of the lawns, the proportion of the area in lawns, or the percentage of lawns that may be treated in a given geographic area. There is limited information on soil types, slopes, watering practices, and other aspects that relate to transport and fate of pesticides. We do know that some homeowners will attempt to control pests with chemicals and that others will not control pests at all or will use non-chemical methods. We would expect that in some areas, few homeowners will use pesticides, but in other areas, a high percentage could. As a result, OPP has insufficient information to develop a scenario or address the extent of pesticide use in a residential area.

It is, however, quite necessary to address the potential that home and garden pesticides may affect T&E species, even in the absence of reliable data. Therefore, I have developed a hypothetical scenario, by adapting an existing scenario, to address pesticide use on home lawns

where it is most likely that residential pesticides will be used outdoors. It is exceedingly important to note that there is no quantitative, scientifically valid support for this modified scenario; rather it is based on my best professional judgement. I do note that the original scenario, based on golf course use, does have a sound technical basis, and the home lawn scenario is effectively the same as the golf course scenario. Three approaches will be used. First, the treatment of fairways, greens, and tees will represent situations where a high proportion of homeowners may use a pesticide. Second, I will use a 10% treatment to represent situations where only some homeowners may use a pesticide. Even if OPP cannot reliably determine the percentage of homeowners using a pesticide in a given area, this will provide two estimates. Third, where the risks from lawn use could exceed our criteria by only a modest amount, I can back-calculate the percentage of land that would need to be treated to exceed our criteria. If a smaller percentage is treated, this would then be below our criteria of concern. The percentage here would be not just of lawns, but of all of the treatable area under consideration; but in urban and highly populated suburban areas, it would be similar to a percentage of lawns. Should reliable data or other information become available, the approach will be altered appropriately.

It is also important to note that pesticides used in urban areas can be expected to transport considerable distances if they should run off on to concrete or asphalt, such as with streets (e.g., TDK Environmental, 2001). This makes any quantitative analysis very difficult to address aquatic exposure from home use. It also indicates that a no-use or no-spray buffer approach for protection, which we consider quite viable for agricultural areas, may not be particularly useful for urban areas.

Finally, the applicability of the overall EEC scenario, i.e., the 10 hectare watershed draining into a one hectare farm pond, may not be appropriate for a number of T&E species living in rivers or lakes. This scenario is intended to provide a “worst-case” assessment of EECs, but very many T&E fish do not live in ponds, and very many T&E fish do not have all of the habitat surrounding their environment treated with a pesticide. OPP does believe that the EECs from the farm pond model do represent first order streams, such as those in headwaters areas (Effland, et al. 1999). In many agricultural areas, those first order streams may be upstream from pesticide use, but in other areas, or for some non-agricultural uses such as forestry, the first order streams may receive pesticide runoff and drift. However, larger streams and lakes will very likely have lower, often considerably lower, concentrations of pesticides due to more dilution by the receiving waters. In addition, where persistence is a factor, streams will tend to carry pesticides away from where they enter into the streams, and the models do not allow for this. The variables in size of streams, rivers, and lakes, along with flow rates in the lentic waters and seasonal variation, are large enough to preclude the development of applicable models to represent the diversity of T&E species’ habitats. We can simply qualitatively note that the farm pond model is expected to overestimate EECs in larger bodies of water.

Indirect Effects - We also attempt to protect listed species from indirect effects of pesticides. We note that there is often not a clear distinction between indirect effects on a listed species and adverse modification of critical habitat (discussed below). By considering indirect effects first, we can provide appropriate protection to listed species even where critical habitat has not been

designated. In the case of fish, the indirect concerns are routinely assessed for food and cover.

The primary indirect effect of concern would be for the food source for listed fish. These are best represented by potential effects on aquatic invertebrates, although aquatic plants or plankton may be relevant food sources for some fish species. However, it is not necessary to protect individual organisms that serve as food for listed fish. Thus, our goal is to ensure that pesticides will not impair populations of these aquatic arthropods. In some cases, listed fish may feed on other fish. Because our criteria for protecting the listed fish species is based upon the most sensitive species of fish tested, then by protecting the listed fish species, we are also protecting the species used as prey.

In general, but with some exceptions, pesticides applied in terrestrial environments will not affect the plant material in the water that provides aquatic cover for listed fish. Application rates for herbicides are intended to be efficacious, but are not intended to be excessive. Because only a portion of the effective application rate of an herbicide applied to land will reach water through runoff or drift, the amount is very likely to be below effect levels for aquatic plants. Some of the applied herbicides will degrade through photolysis, hydrolysis, or other processes. In addition, terrestrial herbicide applications are efficacious in part, due to the fact that the product will tend to stay in contact with the foliage or the roots and/or germinating plant parts, when soil applied. With aquatic exposures resulting from terrestrial applications, the pesticide is not placed in immediate contact with the aquatic plant, but rather reaches the plant indirectly after entering the water and being diluted. Aquatic exposure is likely to be transient in flowing waters. However, because of the exceptions where terrestrially applied herbicides could have effects on aquatic plants, OPP does evaluate the sensitivity of aquatic macrophytes to these herbicides to determine if populations of aquatic macrophytes that would serve as cover for T&E fish would be affected.

For most pesticides applied to terrestrial environment, the effects in water, even lentic water, will be relatively transient. Therefore, it is only with very persistent pesticides that any effects would be expected to last into the year following their application. As a result, and excepting those very persistent pesticides, we would not expect that pesticidal modification of the food and cover aspects of critical habitat would be adverse beyond the year of application. Therefore, if a listed salmon or steelhead is not present during the year of application, there would be no concern. If the listed fish is present during the year of application, the effects on food and cover are considered as indirect effects on the fish, rather than as adverse modification of critical habitat.

Designated Critical Habitat - OPP is also required to consult if a pesticide may adversely modify designated critical habitat. In addition to the indirect effects on the fish, we consider that the use of pesticides on land could have such an effect on the critical habitat of aquatic species in a few circumstances. For example, use of herbicides in riparian areas could affect riparian vegetation, especially woody riparian vegetation, which possibly could be an indirect effect on a listed fish. However, there are very few pesticides that are registered for use on riparian vegetation, and the specific uses that may be of concern have to be analyzed on a pesticide by pesticide basis. In

considering the general effects that could occur and that could be a problem for listed salmonids, the primary concern would be for the destruction of vegetation near the stream, particularly vegetation that provides cover or temperature control, or that contributes woody debris to the aquatic environment. Destruction of low growing herbaceous material would be a concern if that destruction resulted in excessive sediment loads getting into the stream, but such increased sediment loads are insignificant from cultivated fields relative to those resulting from the initial cultivation itself. Increased sediment loads from destruction of vegetation could be a concern in uncultivated areas. Any increased pesticide load as a result of destruction of terrestrial herbaceous vegetation would be considered a direct effect and would be addressed through the modeling of estimated environmental concentrations. Such modeling can and does take into account the presence and nature of riparian vegetation on pesticide transport to a body of water.

Risk Assessment Processes - All of our risk assessment procedures, toxicity test methods, and EEC models have been peer-reviewed by OPP's Science Advisory Panel. The data from toxicity tests and environmental fate and transport studies undergo a stringent review and validation process in accordance with "Standard Evaluation Procedures" published for each type of test. In addition, all test data on toxicity or environmental fate and transport are conducted in accordance with Good Laboratory Practice (GLP) regulations (40 CFR Part 160) at least since the GLPs were promulgated in 1989.

The risk assessment process is described in "Hazard Evaluation Division - Standard Evaluation Procedure - Ecological Risk Assessment" by Urban and Cook (1986) (termed Ecological Risk Assessment SEP below), which has been separately provided to National Marine Fisheries Service staff. Although certain aspects and procedures have been updated throughout the years, the basic process and criteria still apply. In a very brief summary: the toxicity information for various taxonomic groups of species is quantitatively compared with the potential exposure information from the different uses and application rates and methods. A risk quotient of toxicity divided by exposure is developed and compared with criteria of concern. The criteria of concern presented by Urban and Cook (1986) are presented in Table 2.

Table 2. Risk quotient criteria for direct and indirect effects on T&E fish

Test data	Risk quotient	Presumption
Acute LC ₅₀	>0.5	Potentially high acute risk
Acute LC ₅₀	>0.1	Risk that may be mitigated through restricted use classification
Acute LC ₅₀	>0.05	Endangered species may be affected acutely, including sublethal effects

Chronic NOEC	>1	Chronic risk; endangered species may be affected chronically, including reproduction and effects on progeny
Acute invertebrate LC ₅₀ ^a	>0.5	May be indirect effects on T&E fish through food supply reduction
Aquatic plant acute EC ₅₀ ^a	>1 ^b	May be indirect effects on aquatic vegetative cover for T&E fish

a. Indirect effects criteria for T&E species are not in Urban and Cook (1986); they were developed subsequently.

b. This criterion has been changed from our earlier requests. The basis is to bring the endangered species criterion for indirect effects on aquatic plant populations in line with EFED's concern levels for these populations.

The Ecological Risk Assessment SEP (pages 2-6) discusses the quantitative estimates of how the acute toxicity data, in combination with the slope of the dose-response curve, can be used to predict the percentage mortality that would occur at the various risk quotients. The discussion indicates that using a "safety factor" of 10, as applies for restricted use classification, one individual in 30,000,000 exposed to the concentration would be likely to die. Using a "safety factor" of 20, as applies to aquatic T&E species, would exponentially increase the margin of safety. It has been calculated by one pesticide registrant (without sufficient information for OPP to validate that number), that the probability of mortality occurring when the LC50 is 1/20th of the EEC is 2.39×10^{-9} , or less than one individual in ten billion. It should be noted that the discussion (originally part of the 1975 regulations for FIFRA) is based upon slopes of primarily organochlorine pesticides, stated to be 4.5 probits per log cycle at that time. As organochlorine pesticides were phased out, OPP undertook an analysis of more current pesticides based on data reported by Johnson and Finley (1980), and determined that the "typical" slope for aquatic toxicity tests for the "more current" pesticides was 9.95. Because the slopes are based upon logarithmically transformed data, the probability of mortality for a pesticide with a 9.95 slope is again exponentially less than for the originally analyzed slope of 4.5.

The above discussion focuses on mortality from acute toxicity. OPP is concerned about other direct effects as well. For chronic and reproductive effects, our criteria ensures that the EEC is below the no-observed-effect-level, where the "effects" include any observable sublethal effects. Because our EEC values are based upon "worst-case" chemical fate and transport data and a small farm pond scenario, it is rare that a non-target organism would be exposed to such concentrations over a period of time, especially for fish that live in lakes or in streams (best professional judgement). Thus, there is no additional safety factor used for the no-observed-effect-concentration, in contrast to the acute data where a safety factor is warranted because the endpoints are a median probability rather than no effect.

Sublethal Effects - With respect to sublethal effects, Tucker and Leitzke (1979) did an extensive review of existing ecotoxicological data on pesticides. Among their findings was that sublethal effects as reported in the literature did not occur at concentrations below one-fourth to one-sixth of the lethal concentrations, when taking into account the same percentages or numbers affected,

test system, duration, species, and other factors. This was termed the “6x hypothesis”. Their review included cholinesterase inhibition, but was largely oriented towards externally observable parameters such as growth, food consumption, behavioral signs of intoxication, avoidance and repellency, and similar parameters. Even reproductive parameters fit into the hypothesis when the duration of the test was considered. This hypothesis supported the use of lethality tests for use in assessing acute ecotoxicological risk, and the lethality tests are well enough established and understood to provide strong statistical confidence, which can not always be achieved with sublethal effects. By providing an appropriate safety factor, the concentrations found in lethality tests can therefore generally be used to protect from sublethal effects. As discussed earlier, the entire focus of the early-life-stage and life-cycle chronic tests is on sublethal effects.

In recent years, Moore and Waring (1996) challenged Atlantic salmon with diazinon and observed effects on olfaction as relates to reproductive physiology and behavior. Their work indicated that diazinon could have sublethal effects of concern for salmon reproduction. However, the nature of their test system, direct exposure of olfactory rosettes, could not be quantitatively related to exposures in the natural environment. Subsequently, Scholz et al. (2000) conducted a non-reproductive behavioral study using whole Chinook salmon in a model stream system that mimicked a natural exposure that is far more relevant to ecological risk assessment than the system used by Moore and Waring (1996). The Scholz et al. (2000) data indicate potential effects of diazinon on Chinook salmon behavior at very low levels, with statistically significant effects at nominal diazinon exposures of 1 ppb, with apparent, but non-significant effects at 0.1 ppb.

It would appear that the Scholz et al (2000) work contradicts the 6x hypothesis for acute effects. The research design, especially the nature and duration of exposure, of the test system used by Scholz et al (2000), along with a lack of dose-response, precludes comparisons with lethal levels in accordance with the 6x hypothesis as used by Tucker and Leitzke (1979). Nevertheless, it is known that olfaction is an exquisitely sensitive sense. And this sense may be particularly well developed in salmon, as would be consistent with its use by salmon in homing (Hasler and Scholz, 1983). So the contradiction of the 6x hypothesis is not surprising. As a result of these findings, the 6x hypothesis needs to be re-evaluated with respect to olfaction. At the same time, because of the sensitivity of olfaction and because the 6x hypothesis has generally stood the test of time otherwise, it would be premature to abandon the hypothesis for other acute sublethal effects until there are additional data.

2. Description of Linuron:

A. Chemical History: Linuron was first registered as a pesticide in 1966.

B: Chemical Description:

- ☐ Common Name: linuron
- ☐ Chemical Name:
☐ 3-(3,4-dichlorophenyl)-1-methoxy-1-methylurea
- ☐ Chemical Family: Substituted urea
- ☐ Case Number: 0047
- ☐ CAS Registry Number: 330-55-2
- ☐ OPP Chemical Code: 035506
- ☐ Empirical Formula: $C_9H_{10}Cl_2N_2O_2$
- ☐ Molecular Weight: 249.1
- ☐ Trade and Other Names: Linuron®, Linex®, Lorox®,
Lorox-Plus®, Gemeni®, Linuron 4L ®
- ☐ Basic Manufacturer: .E.I. du Pont de Nemours Company, Inc,
Griffin Corporation, and Drexel Chemical
Company

Technical linuron is a odorless, white, crystalline flake or powder. It has a melting point of 86-91°C. It is soluble in water to 81 mg/.L at 25°C. It is slightly soluble in aliphatic hydrocarbons and moderately soluble in ethanol and common aromatic solvents.

C. Chemical Use: The following is based on the currently registered uses of linuron:

- ☐ Type of Agent: Herbicide
- ☐ Mode of Action: Inhibition of Photosynthesis
- ☐ Classification: Non-restricted use herbicide
- ☐ Summary of Sites:
 - ▶ Terrestrial Food/Feed Crops: asparagus, carrots, celery
 - ▶ Terrestrial Food + Feed Crops: corn (sweet and field), parsnips,

potatoes, soybeans, and wheat (winter)

- ▶ Terrestrial Non-Food and Feed Crop: Ornamental bulb production (Calla lily, daffodil, Dutch iris, tulip), non-cropland (roadsides, fencerows, etc), cotton
- ▶ Forestry: hybrid poplar plantations (pulpwood source)

Public Health: None

- ▶ Target Pests:
Preemergence: Florida beggarweed, carpet weed, chickweed, common dayflower, Florida pusskey, galinsoga, lampsquarters, mustards, nettle leaf goosefoot, pigweeds, purslane, wild radish, common ragweed, Pennsylvania smartweed, barnyardgrass, canarygrass, crabgrass, foxtails, goosegrass, fall panicum .

Postemergence: ALL THE ABOVE PLUS; annual morningglory, cocklebur, dogfennel, fiddleneck, groundsel, knawel, prickly sida, sesbania, sicklepod, velvet leaf, wild buckwheat, annual ryegrass, broadleaf signalgrass, rattail fescue, Texas panicum,; NOT galinsoga, chickweed common ragweed, wild radish.

- ❑ Formulation Types Registered:
Technical Grade/Manufacturing-Use Product (MUP) Griffin Linuron
Technical - 95% a.i., Drexel Linuron Technical 1 - 95% a.i., Drexel Linuron Technical 2 - 95% a.i. Drexel Linuron Technical Flake - 95% a.i., DuPont Linuron Technical Flake - 97% a.i.
End-use Product, Drexel Linuron 4L Weed Killer, 41.0% a.i., Drexel Linuron DF, 50% a.i., Griffin Linex 4L Herbicide, 40.6% a.i.
Multiple Ingredient Formulations, 56.9% linuron + chlorimuron, 30.8% linuron + atrazine

- ❑ Methods of Application:

- ▶ Equipment: ground boom, chemigation system
- ▶ Method and Rate: Chemigation (potatoes only), high volume spray (dilute), and low volume spray (concentrate). Maximum use rate for all crops is 1 lb/A.
- ▶ Timing: Linuron products are applied after crop planting, either preemergence or postemergence

❑ Rates of Application (for CA, WA, OR, ID):

- ▶ ASPARAGUS (CA, OR, WA): Direct seeded/newly planted crowns, applied by ground boom: **Preemergence, 1-2 lb a.i./A**, using band of activated charcoal over seed, **postemergence**, 1 or 2 applications at **0.5-1 lb a.i./A**. Established, applied by ground boom. **Preemergence, 1-2 lb a.i./A**. **Postemergence**, 1 to 4 applications at **0.5-1 lb a.i./A**. Do not exceed 4 lb a.i./A per season, PHI = 1 day. **Directed postemergence** application of **2-4 lb a.i./A** may be used to control dudain melon.
- ▶ BULBS (CA), Calla lily, daffodil, Dutch iris, Tulip. After planting, during the growing season, **preemergence 1 lb a.i./A** by ground boom.
- ▶ CARROTS, **postemergence 0.75-1.5 lb a.i./A** by ground boom. If repeat applications are made do not exceed 3 lb a.i./A, PHI = 14 days
- ▶ CELERY, **post-transplant** ground boom application, **0.75-1.5 lb a.i./A** after celery is established but before it is 6 inches tall..
- ▶ PARSNIPS, **preemergence** by ground boom, after planting, before crop emerges, **0.75-1.5 lb a.i./A**, single application
- ▶ WHEAT, WINTER (ID, OR, WA), drill planted. **Preemergence/early post emergence** treatment as soon as possible after planting (though crop may have emerged), by ground spray. West of the Cascades, **1-1.75 lb a.i./A**. East of the Cascades, fall or winter treatment, **0.5-0.75 lb a.i./A**; spring treatment **0.5-0.62 lb a.i./A** as soon as growth starts. Do not apply after the ground has frozen in the fall.
- ▶ NON-CROP SITES (roadsides, fencerows, etc.), **preemergence/early postemergence** treatment, just before weeds emerge or at early seedling stage, **1-3 lb a.i./A** by ground spray.

D. Environmental Fate: Linuron appears to be moderately persistent and relatively immobile. Degradation appears to be primarily microbially mediated, with an aerobic soil half-life ($t_{1/2}$) of 49 days and an anerobic, aqueous $t_{1/2}$ of <21 days. Abiotic processes, such as hydrolysis showed a $t_{1/2}$ >30 days for pH 5, 7, and 9. The registrant calculated an average $t_{1/2}$ of 945 days. At 30 days, 98.4% of the labeled linuron remained. Minor degradates were 3,4-dichlorobenzenamine (DCA), N-(3,4-dichlorophenyl)-N'-methylurea (DCPMU), N-(3,4-dichlorophenyl)-N'-methoxyurea

DML, and (3,4-dichlorophenyl)urea (DCPU). All were present at <1% of the label content.

Aqueous photolysis demonstrated a $t_{1/2}$ >30 days. Registrant calculated $t_{1/2}$ was 45 days. The study was conducted in sterile water, buffered to pH5, under natural light at 25°C. At 30 days post treatment, 61.6% of applied radioactivity remained as parent linuron. Volatiles accounted for 10.% of the label, while 8 unidentified degradates accounted for 5.1%.

The relatively low vapor pressure (1.5×10^{-5} mm Hg at 24°C) suggests that volatility and subsequent photolysis in the atmosphere would not be a major factor in degradation of parent linuron. Data obtained in California indicates $t_{1/2}$'s of from 75 to 100 days in terrestrial field dissipation studies. In Delaware, terrestrial field dissipation $t_{1/2}$ was 57 days.

Under aerobic soil conditions, a $t_{1/2}$ of 49 days was observed. The primary non-volatile degradate was 3-(3,4-dichlorophenyl)-1-methylurea, at 3.0%, 3-(3,4-dichlorophenyl)-1-methoxyurea, 2.1%, and 1-(3,4-dichlorophenyl)urea, at 1.9%.

Anerobic aqueous metabolism of labeled linuron at 24°C demonstrated no linuron remaining at 26 weeks. At three weeks in this system, 10.8% of the label remained as linuron. The major degradates detected were desmethoxy linuron (46.7% at 3 weeks) and desmethoxy monolinuron (75%) at 26 weeks.

Linuron shows limited bioaccumulation in the bluegill sunfish. Bioaccumulation Factors (BCF's) ranged from x40 in muscle, carcass, and whole fish to a maximum BCF of 240 in viscera. 92% of the ^{14}C with which test linuron was labeled was eliminated after the 14 day depuration period.

Linuron is slightly mobile in coarse textured soils ($K_{\text{ads}} = 2.7\text{-}5.0$ ml/g) and relatively immobile in fine-textured soils ($K_{\text{ads}} = 7.51/\text{g}$). A major factor in linuron adsorption is the organic component of the soil. This characteristic may affect the transport of linuron to aquatic environments under some conditions. Movement from surface soils to less biologically active zones will reduce microbial degradation of parent linuron. Adsorption or entrapment in soil particles and dissolved product in surface water may significantly enhance movement following heavy rains.

E. Incidents: A review of linuron incidents conducted by OPP (01/11/01) revealed relatively few incidents, limited to occupational exposures and a single animal incident (pet) since 1992. These incidents were attributed to the known irritating effects of linuron and other substituted urea compounds to the eyes, skin, and mucous membranes. No fish kills were reported.

F. Estimated and actual concentrations of linuron in water: An analysis of toxicity, whether acute or chronic, lethal or sublethal, must be combined with an analysis of how much chemical will be in the water, to determine risks to fish. Risk is a combination of exposure and toxicity. Even a very highly toxic chemical will not pose a risk if there is no exposure, or very

minimal exposure relative to the toxicity. OPP uses a variety of chemical fate and transport data to develop “estimated environmental concentrations” (EECs) from a suite of established models.

The Tier II screening models PRZM and EXAMS with the Index Reservoir (IR) and Percent Crop Area adjustments (IR-PCA PRZM/EXAMS) were used to determine estimated surface water concentration of linuron. The index reservoir represents a potentially vulnerable drinking water source based on the geometry of an actual reservoir and its watershed (located in Illinois). The PCA is a generic watershed based adjustment factor which represents the portion of a watershed planted to a crop and will be applied to pesticide concentration estimates for surface water exposure. The modeled crops include carrot in California. Data used in this assay are shown in Table 3.

Table 3: Data Values Used in the IR-PC PRZM-EXAMS to Determine the Estimated Environmental Concentration for Linuron

Input Variable	Value and Calculation
Crop Evaluated	Carrot
Interval Between Applications (d)	2
Application Efficiency	0.99
Spray Drift Fraction	0.064
Application Method	Ground
DWRATE (day ⁻¹)	0.005
DSRATE (day ⁻¹)	0.005
K _d (mL/mg)	2.7 (sandy loam)
Henry (atm m ² /mole)	6.07 x 10 ⁻⁸ (calculated)
KBACW	0.0003
KBACS (h ⁻¹)	0.0002
KDF (h ⁻¹)	0.0006
KBH, KNH, KAH (h ⁻¹)	stable
KPS (ml/g)	2.7
MWT (g/mole)	249.1

Solubility @ 25°C (ppm)	81
Vapor Pressure (torr)	1.5×10^{-5}

The IR model scenario makes numerous assumptions that may lead to significant over or under estimates of the actual concentration of linuron present in locations that differ significantly because of weather conditions, soil types, water flow rates, and factors beyond the scope of the model. It is not intended that this model is a worst-case scenario. Among the more significant potential variables with regard to linuron, the flow rate in the model is presumed to be constant for the reservoir, while the flow rates in the streams and rivers of concern is clearly seasonal, and highly variable. Natural streams may contain more particulate material than the reservoir and linuron does demonstrate significant adsorption to such material. I am not able to predict if this would increase total linuron content, however a significant change in bioavailability may be present. Seasonal temperature variation is also an uncorrected factor, however the model does not account for stratification and therefore may be more accurate for streams and rivers than for lakes and ponds. With regard to PCA model parameters, it is noted that the estimates are field-level based, and may not accurately reflect pesticide movement in a large, basin scale format. At the present time, however, direct monitoring data are not available for linuron in the area of interest

With these reservations, the calculated EEC for linuron in the California carrot site is shown in Table 4.

Table 4: Estimated Environmental Concentration in Water for Linuron
(Two applications on carrots @ 1lb a.i./A, ground application)

Surface Water Peak (90 th percentile, annual daily maximum)	31.3 µg/L	PCA = 0.87
Surface Water (90 th percentile annual mean)	12.5 µg/L	PCA = 0.87
Surface Water (36-year overall mean)	7.31 µg/L	PCA = 0.87

Direct monitoring data are limited. USGS (1993) sampled a site on the San Joaquin River near Vernakis CA bi-weekly for one year and found a maximum concentration of 0.29 ppb. This result is considered significant because it represents the maximum observed concentration in area that is a major production site for carrots.

In another USGS study, 5196 surface water samples were collected from 40 agricultural streams (nation-wide). Linuron was detected in 2.7% of the samples at a detection limit of 0.01 ppb. The maximum concentration observed was 1.4 ppb.

G. Ecological Effects Toxicity Assessment:

i. Freshwater Fish: The minimum data required to establish the toxicity of linuron technical (for formulation) to freshwater fish is from two species. The preferred species are

rainbow trout (coldwater species) and bluegill sunfish (warm water species). Results of these tests are shown in Table 5.

Table 5: Freshwater Fish, Acute Toxicity

Species	% a.i.	LC ₅₀ (ppm)	Toxicity Class
<i>Oncorhynchus mykiss</i> (trout - static)	96.2	3	Moderately toxic
<i>Lepomis macrochirus</i> (sunfish - static)	96.2	9.6	Moderately toxic

Since the LC₅₀ falls in the range of >1 and <10 ppm, linuron is classified as moderately toxic to freshwater fish. In addition to testing with the technical grade, acute testing with formulated product was required because linuron is registered for use on rights of way, with the potential for aquatic application. Results are shown in Table 6.

Table 6: Freshwater Fish, Acute Toxicity of Lorox 50

Species	% a.i.	LC ₅₀ (ppm)	Toxicity Class
<i>Oncorhynchus mykiss</i> (trout - static)	Lorox 50 (WP)	16.4	Slightly toxic
<i>Lepomis macrochirus</i> (sunfish - static)	Lorox 50 (WP)	16.2	Slightly toxic
<i>Lepomis macrochirus</i> (sunfish - static)	Lorox 50 (DF)	9.2	Moderately toxic

Since the LC₅₀ for the wettable powder was between >10 and <100, it is classified as slightly toxic. The dry flowable product has an LC₅₀ >1 and <10 and is classified as moderately toxic.

ii. Freshwater Fish, Chronic: A freshwater fish early life-cycle test was required for linuron because the exposure may be continuous, recurrent, or multiple, due to multiple applications. The results of this testing are shown in Table 7

Table 7: Chronic Toxicity of Linuron, Early Life Cycle

Species	% a.i.	NOEC (ppm)
<i>Oncorhynchus mykiss</i> (trout - static)	98.4	<0.042

The Maximum Allowable Toxicant Concentration (MATC) could not be determined for linuron because effects on fish length were detected at the lowest test concentration. The agency has requested a core study to determine the No Observable Effects Level (NOEL) for linuron.

iii. Freshwater Invertebrates, Acute: The preferred species for testing linuron toxicity in freshwater invertebrates is the waterflea. Results of acute toxicity tests are shown in Table 8:

Table 8: Acute Toxicity of Linuron in Freshwater Invertebrates.

Species	% a.i.	LC ₅₀ /EC ₅₀ (ppm)	Toxicity Class
<i>Daphnia magna</i> (waterflea)	94.4	0.12	Highly toxic

Since the EC₅₀ is < 1 ppm, linuron is categorized as highly toxic to freshwater invertebrates on an acute basis.

iv. Freshwater Invertebrates, Chronic Toxicity: A freshwater invertebrate, early life - cycle test is required for linuron due to acute toxicity and potential for transport to water. Results of this testing are shown in Table 9.

Table 9: Chronic Toxicity of Linuron to Freshwater Invertebrates

Species	% a.i.	Results
<i>Daphnia magna</i> (waterflea)	98.4	MATC > 0.13 <0.24

Additional testing has been requested by the Agency because the MATC is greater than 0.13 and less than 0.24, which appears inconsistent with the acute data. In addition, it appeared that invertebrates were more sensitive than fish on an acute basis, but appear less sensitive in the chronic tests.

v. Estuarine and Marine Fish, Acute Toxicity: Toxicity testing of linuron in marine/estuarine fish was required. The preferred species is sheepshead minnow. Results of these tests are shown in Table 10.

Table 10: Acute Toxicity of Linuron in Marine/Estuarine Fish

Species	% a.i.	96 hour LC ₅₀ (ppm)	Toxicity Class
<i>Cyprinodon variegatus</i> (sheepshead minnow)	98.4	0.89	Highly toxic

Since the LC₅₀ is <1 and >0.1 ppm, linuron is classified as highly toxic to marine/estuarine fish on an acute basis.

vi. Estuarine/Marine Fish, Chronic Toxicity: Estuarine/marine fish chronic toxicity, early life-cycle testing was required for linuron due to its use for rights of way, which may cross marine/estuarine habitat. Results of this testing were not available as yet.

vii. Estuarine and Marine Invertebrate Acute Toxicity: Testing was performed to determine the acute toxicity of linuron on marine/estuarine invertebrates. The preferred species are mysid shrimp and eastern oyster. Results are shown in Table 11.

Table 11: Acute Toxicity of Linuron to Marine/Estuarine Invertebrates

Species	% a.i.	LC ₅₀ /EC ₅₀ (ppm)	Toxicity Class
<i>Crassostrea virginica</i> (oyster)	98.4	0.89	Highly toxic
<i>Americamysis bahia</i> (mysid shrimp)	98.4	5.4	Moderately toxic

Since the LC₅₀/EC₅₀ of linuron falls in the range of <1 to 10 ppm for mysid shrimp, it is classified as moderately toxic. Linuron is highly toxic to eastern oyster, with an LC₅₀/EC₅₀ between <1 and >0.1.

viii. Estuarine/Marine Invertebrates, Chronic Toxicity: Testing for chronic toxicity of linuron was required, however at the time the RED was published and this review prepared, such data had not been submitted.

The general characterization of linuron toxicity for fresh water and marine/estuarine organisms is that it ranges from slightly toxic to highly toxic. Some distinct inconsistencies in the testing data were observed, particularly between freshwater fish and freshwater invertebrates. A review of the Agency data base (TOXDATA) confirmed considerable variation in quantitative toxicity results, however the general toxicity classifications appeared to be accurate.

H. Risk Quotients for Subject Species:

Based on toxicity and EEC data, risk quotients were calculated relevant to the T&E species of interest in California and Pacific Northwest ESUs. The results of these calculations are presented in Table 12. The EEC (Estimated Environmental Concentration) used to calculate the Risk Quotients (RQs) were derived from two distinct models. One assumed runoff to a 6' pond from the treated crop site (Model A). The second is based on expected runoff into a 6" body of water or wetland (Model B). The B model is used for linuron only for evaluation of use in the treatment of Rights of Way.

Table 12: Risk Quotient Determinations for Freshwater Fish and Invertebrates

Site	Application Rate	Fish (model) ¹	Invertebrate (model) ¹
Carrots, celery, corn, cottonseed, parsley, parsnips, sorghum, ornamentals herbaceous plants	1.5 lb a.i./A	0.021(A)	0.15(A)
Field Corn	1.54 lb a.i./A	0.21(A)	0.157(A)
Winter wheat	1.75 lb a.i./A	0.24(A)	0.178(A)
Potatoes, poplar	2.0 lb a.i./A	0.027(A)	0.203(A)
Soybeans, non-ag, ROW, fence rows, hedgerows, uncult. areas/soils	3.0 lb a.i./A	0.041(A) 0.49(B)	0.305(A) 3.67(B)

Asparagus	4.0 lb a.i./A	0.055(A)	0.4(A)
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¹ A = runoff to 6' pond, B= runoff to 6" wetland

The results indicate the Endangered Species LOC (Level of Concern) was exceeded for all sites in aquatic invertebrates. Endangered Species LOC was exceeded for freshwater fish with field corn, winter wheat, asparagus, and in the wetland runoff model of rights of way treatment (ROW). Restricted Use LOC was exceeded for fish with field corn, wheat, asparagus, and ROW under model B. The Restricted Use LOC was exceeded for invertebrates with potatoes, poplar, soybeans, ROW (etc), and asparagus. The High Risk LOC for invertebrates was exceeded for ROW sites under model B.

I. Discussion and Characterization of Risk Assessment.

Linuron is categorized as being slightly to highly toxic across the spectrum of species tested. It is somewhat persistent in the environment, commonly applied multiple times in the growing season, and demonstrates a modest tendency to bio-accumulate in fish. There are several exceedences for its use, particularly for rights of way and asparagus. Invertebrate High Risk LOC is exceeded for ROW use when linuron runoff is to wetlands. The indirect effects, through loss of food supplies, could be significant since wetlands frequently are breeding areas for many aquatic invertebrates that subsequently may enter streams and rivers.

J. Existing Protections: Curenly the expected precautions regarding spray drift and personal safety measures are components of the label language for linuron. In addition, specific measures are included regarding application rates based on geographic location. These are intended to insure linuron is not applied to sandy soils or to soils with very low organic content.

K. Proposed Protections. Proposed changes include a prohibition against aerial application, restriction on sand and/or loamy/sand soils, and a prohibition agains use on any soil with <1% organic content (sand, limestone, etc.). Application rate reductions for Rights of Way and Hybrid Poplar (midwest only) and rate reduction for soybeans (1..0 lb a.i./A), field corn (0.75 lb a.i./A), potatoes (1.5 lb a.i./A), and asparagus (2.0 lb a.i./A). Applications are to be reduced to 1 per year for field corn, soybeans, and potatoes and 3 per year for asparagus. Warning advisories against application that may contaminate surface water are included in the label language.

In addition, the RED considers making linuron a restricted use pesticide, however at the time of this review the current labels available do not indicate this action has been taken.

3. Description of Pacific salmon and steelhead Evolutionarily Significant Units relative to linuron use sites.

The following review of linuron use in California and the Pacific Northwest is derived from several sources. Califonia data are taken directly from the Department of Pesticide Regulations published census and provides a tabulation of actual chemical use in each county.

The tables for Idaho, Oregon, and Washington are constructed with the 1997 USDA Census of Agriculture as the basis for crops and the estimated use data contained in the RED for linuron. This is identified in the tables as the acres treated. For the northwest states, the amount of chemical applied is calculated on the basis of maximum application rates, annually, identified in the RED and on product labels. Application rates for winter wheat exist, and the Pacific Northwest Weed Management Handbook (2003, Oregon State U., Washington State U., and Idaho State U.) provides use guidelines for wheat East of the Cascades (0.5 lb a.i./A/season) for linuron. However, data from the USGS (1999) analysis of chemicals used indicates that a nationwide total of 1,974,832 lbs a.i. was applied and 100% of this is accounted for in use on crops that do not include wheat. Similar use patterns were identified by the National Center for Food and Agriculture Policy (NCFAP), with an estimate of 516,133 lbs a.i. applied in 1997. EPA use data indicates that 79% of the applied linuron is in soybeans (in the midwest). Other sites, not including wheat, range from <1% to 5% and account for 99% of the total linuron applications. Review of the USGS agricultural map (Attachment 5) confirms the very low levels of linuron use in the northwest, with most areas, including all of Idaho, being reported as having no linuron use. Recent information provided by the Washington State Department of Agriculture indicates that linuron is used as a spot treatment for selected weeds. In 2003 they report a state total of 750 lbs a.i. of linuron was applied to 2,420,000 A of wheat. This, as an average, equals an application rate of 0.0003 lbs a.i./A, a statistically insignificant amount. I have, therefore, made my determinations based on no linuron applications to wheat. For other crops reviewed, the maximum lb a.i./year was used as the rate for determining linuron total pounds applied. It is anticipated that this amount is an overestimate of actual use, however it represents the best available data at the time of review. It is important to note, however, that if linuron is applied to wheat in the future, at already registered application rates (0.75-1.75 lbs a.i./A) the large number of acres planted with winter wheat in Oregon, Idaho, and Washington (>500,000 in some ESUs) could result in very much larger exposure of the T&E species to pesticide and significantly increased risk. The most likely areas at increased risk would be in the Upper and Middle Columbia and Snake River ESU's.

Data are tabulated for tomatoes in California, where Special Local Needs (SLN, FIFRA 24c) registrations CA78016300 and CA79009600 allow application to tomatoes. Alfalfa application is only available in California. No SLNs were identified for Oregon, Idaho, or Washington for tomato application, and cotton is not reported in the census within these states, leading to the conclusion that carrots, asparagus, and winter wheat are the potential registered uses of linuron in these states.

Within California a small amount of the chemical is reported as being applied to sites not currently registered (*ie.* Brussels sprouts, structural pest control, etc.). The California Department of Pesticide Regulation has been notified. The presence of such relatively minor uses is included only because they were reported by California in the pesticide use survey and it contributes, very modestly, to the total amount of linuron potentially affecting the selected ESUs.

1. Southern California Steelhead ESU

The Southern California steelhead ESU was proposed for listing as endangered on August 9, 1996 (61FR41541-41561) and the listing was made final a year later (62FR43937-43954, August 18, 1997). Critical Habitat was proposed February 5, 1999 (64FR5740-5754) and designated on February 16, 2000 (65FR7764-7787). This ESU ranges from the Santa Maria River in San Luis Obispo County south to San Mateo Creek in San Diego County. Steelhead from this ESU may also occur in Santa Barbara, Ventura and Los Angeles counties, but this ESU apparently is no longer considered to be extant in Orange County (65FR79328-79336, December 19, 2000). Hydrologic units in this ESU are Cuyama (upstream barrier - Vaquero Dam), Santa Maria, San Antonio, Santa Ynez (upstream barrier - Bradbury Dam), Santa Barbara Coastal, Ventura (upstream barriers - Casitas Dam, Robles Dam, Matilja Dam, Vern Freeman Diversion Dam), Santa Clara (upstream barrier - Santa Felicia Dam), Calleguas, and Santa Monica Bay (upstream barrier - Rindge Dam). Counties comprising this ESU show a very high percentage of declining and extinct populations.

River entry ranges from early November through June, with peaks in January and February. Spawning primarily begins in January and continues through early June, with peak spawning in February and March.

Within San Diego County, the San Mateo Creek runs through Camp Pendleton Marine Base and into the Cleveland National Forest. While there are agricultural uses of pesticides in other parts of California within the range of this ESU, it would appear that there are no such uses in the vicinity of San Mateo Creek. Within Los Angeles County, this steelhead occurs in Malibu Creek and possibly, but unlikely, Topanga Creek. Neither of these creeks drain agricultural areas and there are no residential uses for this pesticide. There is a potential for steelhead in waters that drain agricultural areas in Ventura, Santa Barbara, and San Luis Obispo counties, but the small quantities of linuron used make effects highly unlikely. Usage of linuron in counties where this ESU occurs are presented in Table 13.

Table 13. Counties supporting the Southern California steelhead ESU

County	Site	Acres Treated	lbs a.i. Applied
Los Angeles	carrot	8468	6200
San Diego	corn, sweet	20	18
San Luis Obispo	carrot	1284	1219
San Luis Obispo	celery	123	519
San Luis Obispo	asparagus	24	31
Santa Barbara	asparagus	581	711
Santa Barbara	carrot	4996	3862

Santa Barbara	celery	2099	1196
Santa Barbara	lettuce ¹	9	5
Santa Barbara	otdr flowers	1	1
Santa Barbara	otdr transplants	46	16
Ventura	carrot	607	486
Ventura	celery	369	2817
Ventura	lettuce ¹	8	4
Ventura	rights of way	NR	25
Ventura	unknown ¹	NR	10

¹ Not a currently registered use.

There is significant use of linuron in the Southern California Steelhead ESU. As a currently unrestricted product, the effects of the dense population centers (Los Angeles, San Diego, Anaheim, etc.) raise additional concerns as to the effects of linuron usage on the endangered steelhead population from non-agricultural uses. The previously noted exceedences of the LOC's for both fish and invertebrates leads me to believe that linuron may have an indirect effect in this ESU, however it is not likely to adversely affect the endangered fish species..

2. South Central California Steelhead ESU

The South Central California steelhead ESU was proposed for listing as endangered on August 9, 1996 (61FR41541-41561) and the listing was made final, as threatened, a year later (62FR43937-43954, August 18, 1997). Critical Habitat was proposed February 5, 1999 (64FR5740-5754) and designated on February 16, 2000 (65FR7764-7787). This coastal steelhead ESU occupies rivers from the Pajaro River, Santa Cruz County, to (but not including) the Santa Maria River, San Luis Obispo County. Most rivers in this ESU drain the Santa Lucia Mountain Range, the southernmost unit of the California Coast Ranges (62FR43937-43954, August 18, 1997). River entry ranges from late November through March, with spawning occurring from January through April.

This ESU includes the Hydrologic units of Pajaro (upstream barriers - Chesbro Reservoir, North Fork Pachero Reservoir), Estrella, Salinas (upstream barriers - Nacimiento Reservoir, Salinas Dam, San Antonio Reservoir), Central Coastal (upstream barriers - Lopez Dam, Whale Rock Reservoir), Alisa-Elkhorn Sloughs, and Carmel. Counties of occurrence include Santa Cruz, San Benito, Monterey, and San Luis Obispo. There are agricultural areas in these counties, and these areas would be drained by waters where steelhead critical habitat occurs.

Table 14: Counties supporting the South Central California steelhead ESU

County	Site	Acres Treated	lbs. a.i. Applied
Monterey	asparagus	755	974
Monterey	beet ¹	1	1
Monterey	carrot	3441	2835
Monterey	cauliflower ¹	8	1
Monterey	celery	847	468
Monterey	mustard ¹	4	4
Monterey	otdr flower	12	10
Monterey	otdr transplant	104	145
Monterey	radish ¹	2	2
Monterey	research	NR	1
Monterey	rights of way	NR	9
San Benito	asparagus	117	91
San Benito	carrot	54	36
San Benito	celery	15	6
San Benito	rangeland	1	2
San Benito	research	NR	2
San Benito	uncultivated ag	1	1
San Mateo	otdr flowers	11	23
San Luis Obispo	asparagus	24	31
San Luis Obispo	carrot	1284	1219
San Luis Obispo	celery	1236	419
Santa Clara			None
Santa Cruz			None

¹ Not a currently registered use.

In the South Central California Steelhead ESU the major sites for linuron are in Monterey and San Luis Obispo Counties. The river sites of concern extend from the Pajaro River in Santa Cruz county to near, but not including, the Santa Maria River in San Luis Obispo county. This suggests

the linuron use in Monterey County (~4,500 lbs) would be of greatest significance. I conclude this may have some effect on the ESU, but is not likely to adversely affect the population due to the large geographic area involved.

3. Central California Coast Steelhead ESU

The Central California coast steelhead ESU was proposed for listing as endangered on August 9, 1996 (61FR41541-41561) and the listing was made final, as threatened, a year later (62FR43937-43954, August 18, 1997). Critical Habitat was proposed February 5, 1999 (64FR5740-5754) and designated on February 16, 2000 (65FR7764-7787). This coastal steelhead ESU occupies California river basins from the Russian River, Sonoma County, to Aptos Creek, Santa Cruz County, (inclusive), and the drainage of San Francisco and San Pablo Bays eastward to the Napa River (inclusive), Napa County. The Sacramento-San Joaquin River Basin of the Central Valley of California is excluded. Steelhead in most tributary streams in San Francisco and San Pablo Bays appear to have been extirpated, whereas most coastal streams sampled in the central California coast region do contain steelhead.

Only winter steelhead are found in this ESU and those to the south. River entry ranges from October in the larger basins, late November in the smaller coastal basins, and continues through June. Steelhead spawning begins in November in the larger basins, December in the smaller coastal basins, and can continue through April with peak spawning generally in February and March. Hydrologic units in this ESU include Russian (upstream barriers - Coyote Dam, Warm Springs Dam), Bodega Bay, Suisun Bay, San Pablo Bay (upstream barriers - Phoenix Dam, San Pablo Dam), Coyote (upstream barriers - Almaden, Anderson, Calero, Guadalupe, Stevens Creek, and Vasona Reservoirs, Searsville Lake), San Francisco Bay (upstream barriers - Calveras Reservoir, Chabot Dam, Crystal Springs Reservoir, Del Valle Reservoir, San Antonio Reservoir), San Francisco Coastal South (upstream barrier - Pilarcitos Dam), and San Lorenzo-Soquel (upstream barrier - Newell Dam).

Counties of occurrence for this ESU are Santa Cruz, San Mateo, San Francisco, Marin, Sonoma, Mendocino, Napa, Alameda, Contra Costa, Solano, and Santa Clara counties. Usage of linuron in the counties where the Central California coast steelhead ESU is shown in Table 15.

Table 15: Counties supporting the Central California Coast steelhead ESU

County	Site	Acres Treated	lbs. a.i. Applied
Alameda			None
Contra Costa			None
Marin			None
Mendocino			None

Napa			None
San Francisco			None
San Mateo	otdr flowers	11	23
Santa Clara			None
Santa Cruz			None
Solano	uncultivated ag	12	12
Sonoma			None

¹ Not a currently registered use.

There is minimal use of linuron in the Central California Coast steelhead ESU (<50 lbs total) and I expect no effects from its use.

4. California Central Valley Steelhead ESU

The California Central Valley steelhead ESU was proposed for listing as endangered on August 9, 1996 (61FR41541-41561) and the listing was made final in 1998 (63FR 13347-13371, March 18, 1998). Critical Habitat was proposed February 5, 1999 (64FR5740-5754) and designated on February 16, 2000 (65FR7764-7787).

This ESU includes populations ranging from Shasta, Trinity, and Whiskeytown areas, along with other Sacramento River tributaries in the North, down the Central Valley along the San Joaquin River to and including the Merced River in the South, and then into San Pablo and San Francisco Bays. Counties at least partly within this area are Alameda, Amador, Butte, Calaveras, Colusa, Contra Costa, Glenn, Marin, Merced, Nevada, Placer, Sacramento, San Francisco, San Joaquin, San Mateo, Solano, Sonoma, Stanislaus, Sutter, Tehama, Tuloumne, Yolo, and Yuba. A large proportion of this area is heavily agricultural. Usage of linuron in counties where the California Central Valley steelhead ESU occurs is presented in Table 16.

Table 16: Counties supporting the California Central Valley steelhead ESU.

County	Site	Acres Treated	lbs. a.i. Applied
Alameda			None
Amador			None
Butte			None
Calaveras			None
Contra Costa			None

Glenn	carrot	4	7
Marin			None
Merced	carrot	9	9
Merced	otdr plants	2	1
Nevada			None
Placer			None
Sacramento			None
San Joaquin	asparagus	2034	1524
San Joaquin	carrot	150	159
San Francisco			None
San Mateo	otdr plants	11	23
Shasta			None
Solano	uncultivated ag	12	12
Sonoma			None
Stanislaus			None
Sutter			None
Tehama			None
Tuolumne			None
Yolo	carrot	76	90
Yuba			None

¹ Not a currently registered use.

The California Central Valley Steelhead ESU is not subjected to heavy linuron application, particularly relative to the large area occupied by it. The only major site is in San Joaquin county. There is no effects on the T&E species under review.

5. Northern California Steelhead ESU

The Northern California steelhead ESU was proposed for listing as threatened on February 11, 2000 (65FR6960-6975) and the listing was made final on June 7, 2000 (65FR36074-36094). Critical Habitat has not yet been officially established.

This Northern California coastal steelhead ESU occupies river basins from Redwood Creek in Humboldt County, CA to the Gualala River, inclusive, in Mendocino County, CA. River entry ranges from August through June and spawning from December through April, with peak spawning in January in the larger basins and in late February and March in the smaller coastal basins. The Northern California ESU has both winter and summer steelhead, including what is presently considered to be the southernmost population of summer steelhead, in the Middle Fork Eel River. Counties included appear to be Humboldt, Mendocino, Trinity, and Lake. Table 17 shows the use of linuron in the counties where the Northern California steelhead ESU occurs.

Table 17: Counties supporting the Northern California steelhead ESU

County	Site	Acres Treated	lbs. a.i. Applied
Humboldt			None
Lake			None
Mendocino			None
Trinity			None

¹ Not a currently registered use.

Linuron is not used in the Northern California Steelhead ESU and will have no effect on the species of concern.

6. Upper Columbia River steelhead ESU

The Upper Columbia River steelhead ESU was proposed for listing as endangered on August 9, 1996 (61FR41541-41561) and the listing was made final a year later (62FR43937-43954, August 18, 1997). Critical Habitat was proposed February 5, 1999 (64FR5740-5754) and designated on February 16, 2000 (65FR7764-7787).

The Upper Columbia River steelhead ESU ranges from several northern rivers close to the Canadian border in central Washington (Okanogan and Chelan counties) to the mouth of the Columbia River. The primary area for spawning and growth through the smolt stage of this ESU is from the Yakima River in south Central Washington upstream. Hydrologic units within the spawning and rearing habitat of the Upper Columbia River steelhead ESU and their upstream barriers are Chief Joseph (upstream barrier - Chief Joseph Dam), Okanogan, Similkameen, Methow, Upper Columbia-Entiat, Wenatchee, Moses-Coulee, and Upper Columbia-Priest Rapids. Within the spawning and rearing areas, counties are Chelan, Douglas, Okanogan, Grant, Benton, Franklin, Kittitas, and Yakima, all in Washington.

Areas downstream from the Yakima River are used for migration. Additional counties through which the ESU migrates are Walla Walla, Klickitat, Skamania, Clark, Columbia, Cowlitz, Wahkiakum, and Pacific, Washington; and Gilliam, Morrow, Sherman, Umatilla, Wasco, Hood River, Multnomah, Columbia, and Clatsop, Oregon.

Tables 18 and 19 show the cropping information and maximum potential linuron use for Washington counties where the Upper Columbia River steelhead ESU is located and for the Oregon and Washington counties where this ESU migrates.

Table 18. Spawning and rearing areas supporting the Upper Columbia River steelhead ESU

State	County	Site	Acres Treated	lbs a.i. Applied
WA	Benton	Asparagus	295	1180
WA	Franklin	Asparagus	1550	6200
WA	Franklin	Carrot	2752	8256
WA	Grant	Asparagus	169	676
WA	Grant	Carrot	1699	5097
WA	Okanogan			Noone
WA	Yakima	Asparagus	1266	5064

Table 19: Oregon and Washington counties that are migration corridors for the Upper Columbia River steelhead ESU.

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Clatsop			None
OR	Columbia			None
OR	Gilliam			None
OR	Hood River			None
OR	Morrow			None
OR	Multnomah			None
OR	Sherman			None
OR	Umatilla	Asparagus	196	784
OR	Wasco			None

WA	Clark			None
WA	Cowlitz			None
WA	Klikitat			None
WA	Pacific			None
WA	Skamannia			None
WA	Wahkiakum			None
WA	Walla Walla	Asparagus	1266	5144

The Upper Columbia River Steelhead ESU courses through major agricultural zones. There is focal, significant, use of linuron in several Washington counties that may indirectly affect the fish in this ESU, however, it is not likely to adversely affect them

7. Snake River Basin steelhead ESU

The Snake River Basin steelhead ESU was proposed for listing as endangered on August 9, 1996 (61FR41541-41561) and the listing was made final a year later (62FR43937-43954, August 18, 1997). Critical Habitat was proposed February 5, 1999 (64FR5740-5754) and designated on February 16, 2000 (65FR7764-7787).

Spawning and early growth areas of this ESU consist of all areas upstream from the confluence of the Snake River and the Columbia River as far as fish passage is possible. Hells Canyon Dam on the Snake River and Dworshak Dam on the Clearwater River, along with Napias Creek Falls near Salmon, Idaho, are named as impassable barriers. These areas include the counties of Wallowa, Baker, Union, and Umatilla (northeastern part) in Oregon; Asotin, Garfield, Columbia, Whitman, Franklin, and Walla Walla in Washington; and Adams, Idaho, Nez Perce, Blaine, Custer, Lemhi, Boise, Valley, Lewis, Clearwater, and Latah in Idaho. Baker County, Oregon, which has a tiny fragment of the Imnaha River watershed was excluded. While a small part of Rock Creek that extends into Baker County, this occurs at 7200 feet in the mountains (partly in a wilderness area) and is of no significance with respect to linuron use in agricultural areas. Similarly excluded are the Upper Grande Ronde watershed tributaries (e.g., Looking Glass and Cabin Creeks) that are barely into higher elevation forested areas of Umatilla County. However, crop areas of Umatilla County are considered in the migratory routes. In Idaho, Blaine and Boise counties technically have waters that are part of the steelhead ESU, but again, these are tiny areas which occur in the Sawtooth National Recreation Area and/or National Forest lands. They have been excluded because they are not relevant to use of linuron. The agricultural areas of Valley County, Idaho, appear to be primarily associated with the Payette River watershed, but there is enough of the Salmon River watershed in this county that it was not able to exclude it.

Critical Habitat also includes the migratory corridors of the Columbia River from the confluence of the Snake River to the Pacific Ocean. Additional counties in the migratory

corridors are Umatilla, Gilliam, Morrow, Sherman, Wasco, Hood River, Multnomah, Columbia, and Clatsop in Oregon; and Benton, Klickitat, Skamania, Clark, Cowlitz, Wahkiakum, and Pacific in Washington.

Tables 20 and 21 show the cropping information for the Pacific Northwest counties where the Snake River Basin steelhead ESU is located and for the Oregon and Washington counties where the fish in this ESU migrate.

Table 20: Rearing/spawning areas supporting the Snake River Basin steelhead ESU .

State	County	Site	Acres Treated	lbs a.i. Applied
ID	Adams			None
ID	Clearwater			None
ID	Custer			None
ID	Idaho			None
ID	Latah			None
ID	Lemhi			None
ID	Nez Perce			None
ID	Valley			None
OR	Union			None
OR	Wallowa			None
WA	Adams	Asparagus	76	304
WA	Asotin			None
WA	Columbia			None
WA	Franklin	Carrot	2752	8256
WA	Franklin	Asparagus	1550	6200
WA	Garfield			None
WA	Walla Walla	Asparagus	255	1020

Table 21. Washington and Oregon counties through which the Snake River Basin steelhead ESU migrates

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Clatsop			None
OR	Columbia			None
OR	Gilliam			None
OR	Hood River			None
OR	Morrow			None
OR	Multnomah			None
OR	Sherman			None
OR	Umatilla	Asparagus	196	784
OR	Wasco			None
WA	Benton			None
WA	Clark			None
WA	Cowlitz			None
WA	Klickitat			None
WA	Wahkiakum			None
WA	Pacific			None
WA	Skamania			None
WA	Walla Walla	Asparagus	255	1020

Application of linuron within the Snake River Basin ESU is modest and I expect no effect the T&E species of concern.

8 Upper Willamette River steelhead ESU

The Upper Willamette River steelhead ESU was proposed for listing as threatened on March 10, 1998 (63FR11798-11809) and the listing was made final a year later (64FR14517-14528, March 25, 1999). Critical Habitat was proposed February 5, 1999 (64FR5740-5754) and designated on February 16, 2000 (65FR7764-7787). Only naturally spawned, winter steelhead trout are included as part of this ESU; where distinguishable, summer-run steelhead trout are not included.

Spawning and rearing areas are river reaches accessible to listed steelhead in the Willamette River and its tributaries above Willamette Falls up through the Calapooia River. This

includes most of Benton, Linn, Polk, Clackamas, Marion, Yamhill, and Washington counties, and small parts of Lincoln and Tillamook counties. However, the latter two counties are small portions in forested areas where linuron would not be used, and these counties are excluded from my analysis. While the Willamette River extends upstream into Lane County, the final Critical Habitat Notice does not include the Willamette River (mainstem, Coastal and Middle forks) in Lane County or the MacKenzie River and other tributaries in this county that were in the proposed Critical Habitat.

Hydrologic units where spawning and rearing occur are Upper Willamette, North Santiam (upstream barrier - Big Cliff Dam), South Santiam (upstream barrier - Green Peter Dam), Middle Willamette, Yamhill, Molalla-Pudding, and Tualatin.

The areas below Willamette Falls and downstream in the Columbia River are considered migration corridors, and include Multnomah, Columbia and Clatsop counties, Oregon, and Clark, Cowlitz, Wahkiakum, and Pacific counties, Washington.

Tables 22 and 23 show the cropping information for Oregon counties where the Upper Willamette River steelhead ESU is located and for the Oregon and Washington counties where this ESU migrates.

Table 22: Spawning and rearing habitat of the Upper Willamette River steelhead ESU.

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Benton			None
OR	Linn			None
OR	Polk			None
OR	Clackamus			None
OR	Marion	Carrot	59	177
OR	Marion			None
OR	Yamhill			None
OR	Washington	Carrot	1	3

Table 23. Oregon and Washington counties that are part of the migration corridors of the Upper Willamette River steelhead ESU.

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Multnomah			None

OR	Clatsop			None
OR	Columbia			None
WA	Clark			None
WA	Cowlitz			None
WA	Wahkiakum			None
WA	Pacific			None

Application of linuron within the Upper Willamette River ESU is quite minimal, and I expect it to have no effect.

9. Lower Columbia River steelhead ESU

The Lower Columbia River steelhead ESU was proposed for listing as endangered on August 9, 1996 (61FR41541-41561) and the listing was made final a year later (62FR43937-43954, August 18, 1997). Critical Habitat was proposed February 5, 1999 (64FR5740-5754) and designated on February 16, 2000 (65FR7764-7787).

This ESU includes all tributaries from the lower Willamette River (below Willamette Falls) to Hood River in Oregon, and from the Cowlitz River up to the Wind River in Washington. These tributaries would provide the spawning and presumably the growth areas for the young steelhead. It is not clear if the young and growing steelhead in the tributaries would use the nearby mainstem of the Columbia prior to downstream migration. If not, the spawning and rearing habitat would occur in the counties of Hood River, Clackamas, and Multnomah counties in Oregon, and Skamania, Clark, and Cowlitz counties in Washington. Tributaries of the extreme lower Columbia River, e.g., Grays River in Pacific and Wahkiakum counties, Washington and John Day River in Clatsop county, Oregon, are not discussed in the Critical Habitat FRNs; because they are not “between” the specified tributaries, they do not appear part of the spawning and rearing habitat for this steelhead ESU. The mainstem of the Columbia River from the mouth to Hood River constitutes the migration corridor. This would additionally include Columbia and Clatsop counties, Oregon, and Pacific and Wahkiakum counties, Washington.

Hydrologic units for this ESU are Middle Columbia-Hood, Lower Columbia-Sandy (upstream barrier - Bull Run Dam 2), Lewis (upstream barrier - Merlin Dam), Lower Columbia-Clatskanie, Lower Cowlitz, Lower Columbia, Clackamas, and Lower Willamette.

Tables 24 and 25 show the cropping information for Oregon and Washington counties where the Lower Columbia River steelhead ESU is located and for the Oregon and Washington counties where this ESU migrates.

Table 24. Spawning/rearing areas for the Lower Columbia steelhead ESU

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Clackamas			None
OR	Hood River			None
OR	Multnomah			None
WA	Clark			None
WA	Cowlitz			None
WA	Skamania			None

*

Table 25: Migratory corridors for the Lower Columbia River Steelhead ESU.

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Clatsop			None
WA	Pacific			None
WA	Wahkiakum			None

Linuron is not used within the Lower Columbia River Steelhead ESU. It will not have an effect the T&E species of interest.

10. Middle Columbia River Steelhead ESU

The Middle Columbia River steelhead ESU was proposed for listing as threatened on March 10, 1998 (63FR11798-11809) and the listing was made final a year later (64FR14517-14528, March 25, 1999). Critical Habitat was proposed February 5, 1999 (64FR5740-5754) and designated on February 16, 2000 (65FR7764-7787).

This steelhead ESU occupies “the Columbia River Basin and tributaries from above the Wind River in Washington and the Hood River in Oregon (exclusive), upstream to, and including, the Yakima River, in Washington.” The Critical Habitat designation indicates the downstream boundary of the ESU to be Mosier Creek in Wasco County, Oregon; this is consistent with Hood River being “excluded” in the listing notice. No downstream boundary is listed for the Washington side of the Columbia River, but if Wind River is part of the Lower Columbia steelhead ESU, it appears that Collins Creek, Skamania County, Washington would be the last stream down river in the Middle Columbia River ESU. Dog Creek may also be part of the ESU, but White Salmon River certainly is, since the Condit Dam is mentioned as an upstream barrier. There is limited data on the status of the Dog and Collins creeks. The only other upstream barrier, in addition to Condit Dam on the White Salmon River is the Pelton Dam on the Deschutes River. As an upstream barrier, this dam would preclude steelhead from reaching the Metolius and

Crooked Rivers as well the upper Deschutes River and its tributaries.

In the John Day River watershed, I have excluded Harney County, Oregon because there is only a tiny amount of the John Day River and several tributary creeks (e.g., Utley, Bear Cougar creeks) which get into high elevation areas (approximately 1700M and higher) of northern Harney County where there are no crops grown. Similarly, the Umatilla River and Walla Walla River get barely into Union County OR, and the Walla Walla River even gets into a tiny piece of Wallowa County, Oregon. But again, these are high elevation areas where crops are not grown, and are excluded counties for this analysis.

The Oregon counties then that appear to have spawning and rearing habitat are Gilliam, Morrow, Umatilla, Sherman, Wasco, Crook, Grant, Wheeler, and Jefferson counties. Hood River, Multnomah, Columbia, and Clatsop counties in Oregon provide migratory habitat. Washington counties providing spawning and rearing habitat would be Benton, Columbia, Franklin, Kittitas, Klickitat, Skamania, Walla Walla, and Yakima, although only a small portion of Franklin County between the Snake River and the Yakima River is included in this ESU. Skamania, Clark, Cowlitz, Wahkiakum, and Pacific Counties in Washington provide migratory corridors.

Tables 26 and 27 show the cropping information for Oregon and Washington counties where the Middle Columbia River steelhead ESU is located and for the Oregon and Washington counties where the fish migrate.

Table 26. Spawning/Rearing areas for the Middle Columbia Steelhead ESU

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Crook	Winter Wheat		None
OR	Gilliam	Winter Wheat		None
OR	Jefferson	Winter Wheat		None
OR	Morrow	Winter Wheat		None
OR	Sherman	Winter Wheat		None
OR	Umatilla	Asparagus	196	784
OR	Wasco			None
OR	Wheeler			None
WA	Benton	Asparagus	295	1180
WA	Columbia			None
WA	Franklin	Asparagus	1550	6200

WA	Franklin	Carrot	2752	8256
WA	Grant	Asparagus	169	676
WA	Grant	Carrot	1699	5097
WA	Grant			None
WA	Kittitat			None
WA	Skamania			None
WA	Walla Walla	Asparagus	255	1020
WA	Yakima	Asparagus	1266	5064

Table 27. Washington and Oregon counties through which the Middle Columbia River steelhead ESU migrates

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Clatsop			None
OR	Columbia			None
OR	Hood River			None
OR	Multnomah			None
WA	Clark			None
WA	Cowlitz			None
WA	Pacific			None
WA	Wakiakum			None

There is focal high use of linuron within the Middle Columbia Steelhead ESU. Of particular concern is the presence of the pesticide within the spawning and rearing areas. The EFED risk analysis demonstrated several exceedences, particularly to macroinvertebrates, a primary food source for young fish. These observations and the relative persistence of linuron lead me to believe it may affect this ESU, through indirect food loss, but the large geographic size of the ESU suggests linuron is not likely to have an adverse effect on this ESU.

B. Chinook salmon

Chinook salmon (*Oncorhynchus tshawytscha*) is the largest salmon species; adults weighing over 120 pounds have been caught in North American waters. Like other Pacific salmon, chinook salmon are anadromous and die after spawning.

Juvenile stream and ocean type chinook salmon have adapted to different ecological niches. Ocean-type chinook salmon, commonly found in coastal streams, tend to utilize estuaries and coastal areas more extensively for juvenile rearing. They typically migrate to sea within the first three months of emergence and spend their ocean life in coastal waters. Summer and fall runs predominate for ocean-type chinook. Stream-type chinook are found most commonly in headwater streams and are much more dependent on freshwater stream ecosystems because of their extended residence in these areas. They often have extensive offshore migrations before returning to their natal streams in the spring or summer months. Stream-type smolts are much larger than their younger ocean-type counterparts and are therefore able to move offshore relatively quickly.

Coast-wide, chinook salmon typically remain at sea for 2 to 4 years, with the exception of a small proportion of yearling males (called jack salmon) which mature in freshwater or return after 2 or 3 months in salt water. Ocean-type chinook salmon tend to migrate along the coast, while stream-type chinook salmon are found far from the coast in the central North Pacific. They return to their natal streams with a high degree of fidelity. Seasonal “runs” (i.e., spring, summer, fall, or winter), which may be related to local temperature and water flow regimes, have been identified on the basis of when adult chinook salmon enter freshwater to begin their spawning migration. Egg deposition must occur at a time to ensure that fry emerge during the following spring when the river or estuarine productivity is sufficient for juvenile survival and growth.

Adult female chinook will prepare a spawning bed, called a redds, in a stream area with suitable gravel composition, water depth and velocity. After laying eggs in a redds, adult chinook will guard the redds from 4 to 25 days before dying. Chinook salmon eggs will hatch, depending upon water temperatures, between 90 to 150 days after deposition. Juvenile chinook may spend from 3 months to 2 years in freshwater after emergence and before migrating to estuarine areas as smolts, and then into the ocean to feed and mature. Historically, chinook salmon ranged as far south as the Ventura River, California, and their northern extent reaches the Russian Far East.

1. Sacramento River Winter-run Chinook Salmon ESU

The Sacramento River Winter-run chinook was emergency listed as threatened with critical habitat designated in 1989 (54FR32085-32088, August 4, 1989). This emergency listing provided interim protection and was followed by (1) a proposed rule to list the winter-run on March 20, 1990, (2) a second emergency rule on April 20, 1990, and (3) a formal listing on November 20, 1990 (59FR440-441, January 4, 1994). A somewhat expanded critical habitat was proposed in 1992 (57FR36626-36632, August 14, 1992) and made final in 1993 (58FR33212-33219, June 16, 1993). In 1994, the winter-run was reclassified as endangered because of significant declines and continued threats (59FR440-441, January 4, 1994).

Critical Habitat has been designated to include the Sacramento River from Keswick Dam, Shasta County (river mile 302) to Chipps Island (river mile 0) at the west end of the Sacramento-San Joaquin delta, and then westward through most of the fresh or estuarine waters, north of the Oakland Bay Bridge, to the ocean. Estuarine sloughs in San Pablo and San Francisco bays are

excluded (58FR33212-33219, June 16, 1993).

Table 28 shows the linuron usage in California counties supporting the Sacramento River winter-run chinook salmon ESU. Spawning areas are primarily in Shasta and Tehama counties above the Red Bluff diversion dam.

Table 28: California counties supporting the Sacramento River, winter-run chinook ESU.

County	Site	Acres Treated	lbs a.i. Applied
Alameda			None
Butte			None
Contra Costa			None
Glenn	carrot	4	7
Marin			None
Sacramento			None
San Francisco			None
San Mateo			None
Shasta			None
Solano	uncultivated ag	12	12
Sonoma			None
Sutter			None
Tehama			None
Yolo	carrot	76	90
Yolo			None

¹ Not a currently registered use.

Application of linuron within the Sacramento River, winter run, Chinook ESU is reported as a total of 109 pounds, a minimal amount relative to the land mass involved. Its use will not have an effect the T&E species of interest.

2. Snake River Fall-run Chinook Salmon ESU

The Snake River fall-run chinook salmon ESU was proposed as threatened in 1991 (56FR29547-29552, June 27, 1991) and listed about a year later (57FR14653-14663, April 22,

1992). Critical habitat was designated on December 28, 1993 (58FR68543-68554) to include all tributaries of the Snake and Salmon Rivers accessible to Snake River fall-run chinook salmon, except reaches above impassable natural falls and Dworshak and Hells Canyon Dams. The Clearwater River and Palouse River watersheds are included for the fall-run ESU, but not for the spring/summer run. This chinook ESU was proposed for reclassification on December 28, 1994 (59FR66784-57403) as endangered because of critically low levels, based on very sparse runs. However, because of increased runs in the subsequent year, this proposed reclassification was withdrawn (63FR1807-1811, January 12, 1998).

In 1998, NMFS proposed to revise the Snake River fall-run chinook to include those stocks using the Deschutes River (63FR11482-11520, March 9, 1998). The John Day, Umatilla, and Walla Walla Rivers would be included; however, fall-run chinook in these rivers are believed to have been extirpated. It appears that this proposal has yet to be finalized. I have not included these counties here; however, I would note that the Middle Columbia River steelhead ESU encompasses these basins, and crop information is presented in that section of this analysis.

Hydrologic units with spawning and rearing habitat for this fall-run chinook are the Clearwater, Hells Canyon, Imnaha, Lower Grande Ronde, Lower North Fork Clearwater, Lower Salmon, Lower Snake-Asotin, Lower Snake-Tucannon, and Palouse. These units are in Baker, Umatilla, Wallowa, and Union counties in Oregon; Adams, Asotin, Columbia, Franklin, Garfield, Lincoln, Spokane, Walla Walla, and Whitman counties in Washington; and Adams, Benewah, Clearwater, Idaho, Latah, Lewis, Nez Perce, Shoshone, and Valley counties in Idaho. Custer and Lemhi counties in Idaho are not listed as part of the fall-run ESU, although they are included for the spring/summer-run ESU. Because only high elevation forested areas of Baker and Umatilla counties in Oregon are in the spawning and rearing areas for this fall-run chinook, they were excluded them from consideration because linuron would not be used in these areas.

Table 29 show the cropping information for Pacific Northwest counties where the Snake River fall-run chinook salmon ESU is located. Migration corridors are the same as those in Table 21.

Table 29 : Spawning/rearing areas supporting the Snake River Fall-run chinook salmon ESU

State	County	Site	Acres Treated	lbs a.i. Applied
ID	Adams			None
ID	Benewah			None
ID	Clearwater			None
ID	Idaho			None
ID	Latah			None

ID	Lewis			None
ID	Nez Perce			None
ID	Shoshone			None
OR	Union			None
OR	Wallowa			None
WA	Adams	Asparagus	76	304
WA	Asotin			None
WA	Franklin	Asparagus	1550	6200
WA	Franklin	Carrot	2752	8256
WA	Garfield			None
WA	Walla Walla	Asparagus	255	1020

There is some use of linuron within the Snake River fall run Chinook Salmon ESU, within the spawning and rearing areas, and I anticipate it may indirectly affect the T&E species of concern since its major toxicity is to the food source of young fish, but it is not likely to adversely affect the ESU.

3. Snake River Spring/Summer-run Chinook Salmon

The Snake River Spring/Summer-run chinook salmon ESU was proposed as threatened in 1991 (56FR29542-29547, June 27, 1991) and listed about a year later (57FR14653-14663, April 22, 1992). Critical habitat was designated on December 28, 1993 (58FR68543-68554) to include

all tributaries of the Snake and Salmon Rivers (except the Clearwater River) accessible to Snake River spring/summer chinook salmon. Like the fall-run chinook, the spring/summer-run chinook ESU was proposed for reclassification on December 28, 1994 (59FR66784-57403) as endangered because of critically low levels, based on very sparse runs. However, because of increased runs in subsequent year, this proposed reclassification was withdrawn (63FR1807-1811, January 12, 1998).

Hydrologic units in the potential spawning and rearing areas include Hells Canyon, Imnaha, Lemhi, Little Salmon, Lower Grande Ronde, Lower Middle Fork Salmon, Lower Salmon, Lower Snake-Asotin, Lower Snake-Tucannon, Middle Salmon-Chamberlain, Middle Salmon - Panther, Pahsimerol, South Fork Salmon, Upper Middle Fork Salmon, Upper Grande Ronde, Upper Salmon, and Wallowa. Areas above Hells Canyon Dam are excluded, along with unnamed “impassable natural falls”. Napias Creek Falls, near Salmon, Idaho, was later named an upstream barrier (64FR57399-57403, October 25, 1999). The Grande Ronde, Imnaha, Salmon, and Tucannon subbasins, and Asotin, Granite, and Sheep Creeks were specifically named in the Critical Habitat Notice.

Spawning and rearing counties mentioned in the Critical Habitat Notice include Union, Umatilla, Wallowa, and Baker counties in Oregon; Adams, Blaine, Custer, Idaho, Lemhi, Lewis, Nez Perce, and Valley counties in Idaho; and Asotin, Columbia, Franklin, Garfield, Walla Walla, and Whitman counties in Washington. However, Umatilla and Baker counties in Oregon and Blaine County in Idaho are excluded because accessible river reaches are all well above areas where linuron can be used. Counties with migratory corridors are all of those down stream from the confluence of the Snake and Columbia Rivers.

Table 30 shows the counties where the Snake River spring/summer-run chinook salmon ESU occurs. The cropping information for the migratory corridors is the same as for the Snake River fall-run chinook salmon and is in the Table 21.

Table 30: Spawning/rearing area supporting the Snake River spring/summer chinook ESU

State	County	Site	Acres Treated	lbs a.i. Applied
ID	Adams			None
ID	Idaho			None
ID	Latah			None
ID	Lewis			None
ID	Nez Perce			None
ID	Shoshone			None
ID	Valley			None

OR	Union			None
OR	Wallowa			None
WA	Asotin			None
WA	Franklin	Asparagus	1550	6200
WA	Franklin	Carrot	2752	8256
WA	Garfield			None
WA	Walla Walla	Asparagus	255	1020

There is modest use of linuron within the Snake River spring/summer run Chinook Salmon ESU, within the spawning and rearing areas and this may indirectly affect the T&E species of concern since its major toxicity is to the food source of young fish, but is not likely to adversely affect the ESU.

4. Central Valley Spring-run Chinook Salmon ESU

The Central valley Spring-run chinook salmon ESU was proposed as threatened in 1998 (63FR11482-11520, March 9, 1998) and listed on September 16, 1999 (64FR50393-50415). Critical habitat was designated February 16, 2000 (65FR7764-7787) to encompass all river reaches accessible to listed chinook salmon in the Sacramento River and its tributaries in California, along with the down stream river reaches into San Francisco Bay, north of the Oakland Bay Bridge, and to the Golden Gate Bridge

Hydrologic units and upstream barriers within this ESU are the Sacramento-Lower Cow-Lower Clear, Lower Cottonwood, Sacramento-Lower Thomas (upstream barrier - Black Butte Dam), Sacramento-Stone Corral, Lower Butte (upstream barrier - Chesterville Dam), Lower Feather (upstream barrier - Orville Dam), Lower Yuba, Lower Bear (upstream barrier - Camp Far West Dam), Lower Sacramento, Sacramento-Upper Clear (upstream barriers - Keswick Dam, Whiskey town dam), Upper Elder-Upper Thomas, Upper Cow-Battle, Mill-Big Chico, Upper Butte, Upper Yuba (upstream barrier - Englebright Dam), Suisin Bay, San Pablo Bay, and San Francisco Bay. These areas are said to be in the counties of Shasta, Tehama, Butte, Glenn, Colusa, Sutter, Yolo, Yuba, Placer, Sacramento, Solano, Nevada, Contra Costa, Napa, Alameda, Marin, Sonoma, San Mateo, and San Francisco. I note, however, with San Mateo County being well south of the Oakland Bay Bridge, it is difficult to see why this county was included.

Table 31: California counties supporting the Central Valley spring-run chinook salmon ESU.

County	Site	Acres Treated	Lbs a.i. Applied
Alameda			None

Butte			None
Calaveras			None
Colusa	carrot	131	98
Contra Costa			None
Glenn			None
Merced	carrot	9	9
Merced	otdr plants	2	1
Marin			None
Placer			None
Sacramento			None
San Francisco			None
San Mateo	otdr plants	11	23
Shasta			None
Solano	uncultivated ag	12	12
Sonoma			None
Sutter			None
Tehama			None
Yolo	carrot	76	90
Yuba			None

¹ Not a currently registered use.

Application of linuron within the California Central Valley, spring-run, Chinook ESU is quite low, relative to the land mass involved. Its use will have no effect on the T&E species of interest.

5. California Coastal Chinook Salmon ESU

The California coastal chinook salmon ESU was proposed as threatened in 1998 (63FR11482-11520, March 9, 1998) and listed on September 16, 1999 (64FR50393-50415). Critical habitat was designated February 16, 2000 (65FR7764-7787) to encompass all river reaches and estuarine areas accessible to listed chinook salmon from Redwood Creek (Humboldt County, California) to the Russian River (Sonoma County, California), inclusive.

The Hydrologic units and upstream barriers are Mad-Redwood, Upper Eel (upstream barrier - Scott Dam), Middle Fort Eel, Lower Eel, South Fork Eel, Mattole, Big-Navarro-Garcia, Gualala-Salmon, Russian (upstream barriers - Coyote Dam; Warm Springs Dam), and Bodega Bay. Counties with agricultural areas where linuron could be used are Humboldt, Trinity, Mendocino, Lake, Sonoma, and Marin. A small portion of Glenn County is also included in the Critical Habitat, but linuron would not be used in the forested upper elevation areas.

Table 32: California counties supporting the California coastal chinook salmon ESU.

County	Site	Acres Treated	Lbs a.i. Applied
Humboldt			None
Lake			None
Marin			None
Mendocino			None
Sonoma			None
Trinity			None

¹ Not a currently registered use.

There is no use of linuron within the California Coastal Chinook Salmon ESU, and it will have no effect on the T&E species of interest.

6. Puget Sound Chinook Salmon ESU

The Puget Sound chinook salmon ESU was proposed as threatened in 1998 (63FR11482-11520, March 9, 1998) and listed a year later (64FR14308-14328, March 24, 1999). Critical habitat was designated February 16, 2000 (65FR7764-7787) to encompass all marine, estuarine, and river reaches accessible to listed chinook salmon in Puget Sound and its tributaries, extending out to the Pacific Ocean.

The Hydrologic units and upstream barriers are the Strait of Georgia, San Juan Islands, Nooksack, Upper Skagit, Sauk, Lower Skagit, Stillaguamish, Skykomish, Snoqualmie (upstream barrier - Tolt Dam), Snohomish, Lake Washington (upstream barrier - Landsburg Diversion), Duwamish, Puyallup, Nisqually (upstream barrier - Alder Dam), Deschutes, Skokomish, Hood Canal, Puget Sound, Dungeness-Elwha (upstream barrier - Elwha Dam). Affected counties in Washington, apparently all of which could have spawning and rearing habitat, are Skagit, Whatcom, San Juan, Island, Snohomish, King, Pierce, Thurston, Lewis, Grays Harbor, Mason, Clallam, Jefferson, and Kitsap.

Table 33: Washington counties where the Puget Sound chinook salmon ESU is located.

State	County	Site	Acres Treated	lbs a.i. Applied
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WA	Clallum			None
WA	Grays Harbor			None
WA	Jefferson			None
WA	King	Carrot	8	24
WA	Kitsap	Carrot	1	3
WA	Lewis			None
WA	Mason			None
WA	Pierce			None
WA	San Juan	Carrot	1	3
WA	Skagit	Carrot	389	1167
WA	Snohomish	Carrot	2	6
WA	Thurston			None
WA	Whatcom			None

The Puget Sound Chinook Salmon ESU is located in a body of water that is largely closed except for the northern portion, which opens to the Straits of Juan de Fuca and the Georgia Straits of Canada. The unusual character of Puget Sound results in exceptional tidal activity (greater than ± 20 feet in the Spring) and considerable water movement. This is reflected in the flow patterns of the tributaries serving as salmon habitat. The high rates of water flow, tidal disturbances, and similar factors will greatly enhance movement of linuron and can be expected to quickly reduce its concentration below toxic levels. I anticipate no effects from linuron use on the species of interest in this review.

7. Lower Columbia River Chinook Salmon ESU

The Lower Columbia River chinook salmon ESU was proposed as threatened in 1998 (63FR11482-11520, March 9, 1998) and listed a year later (64FR14308-14328, March 24, 1999). Critical habitat was designated February 16, 2000 (65FR7764-7787) to encompass all river reaches accessible to listed chinook salmon in Columbia River tributaries between the Grays and White Salmon Rivers in Washington and the Willamette and Hood Rivers in Oregon, inclusive, along with the lower Columbia River reaches to the Pacific Ocean.

The Hydrologic units and upstream barriers are the Middle Columbia-Hood (upstream barriers - Condit Dam, The Dalles Dam), Lower Columbia-Sandy (upstream barrier - Bull Run Dam 2), Lewis (upstream barrier - Merlin Dam), Lower Columbia-Clatskanie, Upper Cowlitz, Lower Cowlitz, Lower Columbia, Clackamas, and the Lower Willamette. Spawning and rearing

habitat would be in the counties of Hood River, Waco, Columbia, Clackamas, Marion, Multnomah, and Washington in Oregon, and Klickitat, Skamania, Clark, Cowlitz, Lewis, Wahkiakum, Pacific, Yakima, and Pierce in Washington. Clatsop County appears to be the only county in the critical habitat that does not contain spawning and rearing habitat, although there is only a small part of Marion County that is included as critical habitat. Pierce County, Washington was excluded because the very small part of the Cowlitz River watershed in this county is at a high elevation where linuron would not be used.

Table 34: Oregon and Washington counties where the Lower Columbia River chinook salmon ESU occurs.

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Clackamas			None
OR	Clatsop			None
OR	Hood River			None
OR	Marion	Carrot	59	177
OR	Multnomah			None
OR	Wasco			None
OR	Washington	Carrot	1	3
WA	Clark			None
WA	Cowlitz			None
WA	Klickitat			None
WA	Lewis			None
WA	Pacific			None
WA	Skamania			None
WA	Wakiakum			None

Application of linuron within the Lower Columbia River ESU is quite low and it will have no effect on the species under review.

8. Upper Willamette River Chinook Salmon ESU

The Upper Willamette River Chinook Salmon ESU was proposed as threatened in 1998 (63FR11482-11520, March 9, 1998) and listed a year later (64FR14308-14328, March 24, 1999). Critical habitat was designated February 16, 2000 (65FR7764-7787) to encompass all river

reaches accessible to listed chinook salmon in the Clackamas River and the Willamette River and its tributaries above Willamette Falls, in addition to all down stream river reaches of the Willamette and Columbia Rivers to the Pacific Ocean.

The Hydrologic units included are the Lower Columbia-Sandy, Lower Columbia-Clatskanie, Lower Columbia, Middle Fork Willamette, Coast Fork Willamette (upstream barriers - Cottage Grove Dam, Dorena Dam), Upper Willamette (upstream barrier - Fern Ridge Dam), McKenzie (upstream barrier - Blue River Dam), North Santiam (upstream barrier - Big Cliff Dam), South Santiam (upstream barrier - Green Peter Dam), Middle Willamette, Yamhill, Molalla-Pudding, Tualatin, Clackamas, and Lower Willamette. Spawning and rearing habitat is in the Oregon counties of Clackamas, Douglas, Lane, Benton, Lincoln, Linn, Polk, Marion, Yamhill, Washington, and Tillamook. However, Lincoln and Tillamook counties include salmon habitat only in the forested parts of the coast range where linuron would not be used. Salmon habitat for this ESU is exceedingly limited in Douglas County also, but we cannot rule out future Linuron use in Douglas County.

Tables 35 and 36 show the cropping information for Oregon counties where the Upper Willamette River chinook salmon ESU occurs and for the Oregon and Washington counties where the ESU fish migrate.

Table 35: Spawning/Rearing areas for the Upper Willamette River chinook ESU

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Benton			None
OR	Clackamus			None
OR	Douglas			None
OR	Lane	Carrot	208	624
OR	Linn			None
OR	Marion	Carrot	59	177
OR	Polk			None
OR	Wasco			None
OR	Washington			None
OR	Yamhill			None

Table 36: Migration corridors of the Upper Willamette River chinook salmon ESU.

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Clatsop			None

OR	Columbia			None
OR	Multnomah			None
WA	Clark			None
WA	Cowlitz			None
WA	Pacific			None

Application of linuron within the Upper Willamette River Chinook Salmon ESU is low in the migratory pathways and rearing areas. I expect no effects from its use.

9. Upper Columbia River Spring-run Chinook Salmon ESU

The Upper Columbia River Spring-run Chinook Salmon ESU was proposed as endangered in 1998 (63FR11482-11520, March 9, 1998) and listed a year later (64FR14308-14328, March 24, 1999). Critical habitat was designated February 16, 2000 (65FR7764-7787) to encompass all river reaches accessible to listed chinook salmon in Columbia River tributaries upstream of the Rock Island Dam and downstream of Chief Joseph Dam in Washington, excluding the Okanogan River, as well as all down stream migratory corridors to the Pacific Ocean. Hydrologic units and their upstream barriers are Chief Joseph (Chief Joseph Dam), Similkameen, Methow, Upper Columbia-Entiat, Wenatchee, Upper Columbia-Priest Rapids, Middle Columbia-Lake Wallula, Middle Columbia-Hood, Lower Columbia-Sandy, Lower Columbia-Clatskanie, Lower Columbia, and Lower Willamette. Counties in which spawning and rearing occur are Chelan, Douglas, Okanogan, Grant, Kittitas, and Benton (Table 36), with the lower river reaches being migratory corridors (Table 37).

Most linuron usage occurs upstream from the confluence of the Snake River with the Columbia River, but not as far north as Chelan, and Okanogan counties, where there is limited acreage of potato, the only crop for linuron. However, a modest amount is used on potato below that confluence in counties on either side of the Columbia River, but all upstream of the John Day Dam.

Tables 37 and 38 show the cropping information for Washington counties that support the Upper Columbia River chinook salmon ESU and for the Oregon and Washington counties where the ESU species migrate.

Table 37. Counties Supporting the Upper Columbia Chinook ESU Spawning/Rearing Area

State	County	Site	Acres Treated	lbs a.i. Applied
WA	Benton	Asparagus	295	1180

WA	Chelan			None
WA	Douglas			None
WA	Grant	Asparagus	2	1
WA	Grant	Carrot	1699	5097
WA	Kittitas			None
WA	Okanogan			None
WA	Skamania			None

Table 38: .Migration corridors for the Upper Columbia River Chinook salmon ESU.

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Clatsop			None
OR	Columbia			None
OR	Gilliam			None
OR	Hood River			None
OR	Morrow			None
OR	Multnomah			None
OR	Sherman			None
OR	Umatilla	Asparagus	196	784
OR	Wasco			None
WA	Cowlitz			None
WA	Franklin	Asparagus	1550	6200
WA	Franklin	Carrot	2752	8256
WA	Klckitat			None
WA	Skamania			None
WA	Pacific			None
WA	Walla Walla	Asparagus	255	1020

WA	Yakima	Asparagus	1266	5064
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The Upper Columbia River Chinook ESU courses through major agricultural zones with moderate use of linuron. This suggests that the ESU migratory pathways and rearing areas, may be indirectly affected through a reduction in food sources, but not likely to be adversely Affected.

C. Coho Salmon

Coho salmon, *Oncorhynchus kisutch*, were historically distributed throughout the North Pacific Ocean from central California to Point Hope, AK, through the Aleutian Islands into Asia. Historically, this species probably inhabited most coastal streams in Washington, Oregon, and central and northern California. Some populations may once have migrated hundreds of miles inland to spawn in tributaries of the upper Columbia River in Washington and the Snake River in Idaho.

Coho salmon generally exhibit a relatively simple, 3 year life cycle. Adults typically begin their freshwater spawning migration in the late summer and fall, spawn by mid-winter, then die. Southern populations are somewhat later and spend much less time in the river prior to spawning than do northern coho. Homing fidelity in coho salmon is generally strong; however, their small tributary habitats experience relatively frequent, temporary blockages, and there are a number of examples in which coho salmon have rapidly re-colonized vacant habitat that had only recently become accessible to anadromous fish.

After spawning in late fall and early winter, eggs incubate in redds for 1.5 to 4 months, depending upon the temperature, before hatching as alevins. Following yolk sac absorption, alevins emerge and begin actively feeding as fry. Juveniles rear in fresh water for up to 15 months, then migrate to the ocean as “smolts” in the spring. Coho salmon typically spend two

growing seasons in the ocean before returning to their natal stream. They are most frequently recovered from ocean waters in the vicinity of their spawning streams, with a minority being recovered at adjacent coastal areas, decreasing in number with distance from the natal streams. However, those coho released from Puget Sound, Hood Canal, and the Strait of Juan de Fuca are caught at high levels in Puget Sound, an area not entered by coho salmon from other areas.

1. Central California Coast Coho Salmon ESU

The Central California Coast Coho Salmon ESU includes all coho naturally reproduced in streams between Punta Gorda, Humboldt County, CA and San Lorenzo River, Santa Cruz County, CA, inclusive. This ESU was proposed in 1995 (60FR38011-38030, July 25, 1995) and listed as threatened, with critical habitat designated, on May 5, 1999 (64FR24049-24062). Critical habitat consists of accessible reaches along the coast, including Arroyo Corte Madera

Del Presidio and Corte Madera Creek, tributaries to San Francisco Bay.

Hydrologic units within the boundaries of this ESU are: San Lorenzo-Soquel (upstream barrier - Newell Dam), San Francisco Coastal South, San Pablo Bay (upstream barrier - Phoenix Dam- Phoenix Lake), Tomales-Drake Bays (upstream barriers - Peters Dam-Kent Lake; Seeger Dam-Nicasio Reservoir), Bodega Bay, Russian (upstream barriers - Warm springs dam-Lake Sonoma; Coyote Dam-Lake Mendocino), Gualala-Salmon, and Big-Navarro-Garcia. California counties included are Santa Cruz, San Mateo, Marin, Napa, Sonoma, and Mendocino.

Table 39: California counties supporting the Central California coast Coho salmon ESU.

County	Site	Acres Treated	Lbs a.i. Applied
Marin			None
Mendocino			None
Napa			None
San Mateo	otdr flowers	11	23
Santa Cruz			None
Sonoma			None

Linuron is minimally used within the Central California coast Coho salmon ESU and will, therefore, have no effects on the species of concern.

2. Southern Oregon/Northern California Coast Coho Salmon ESU

The Southern Oregon/Northern California coastal coho salmon ESU was proposed as threatened in 1995 (60FR38011-38030, July 25, 1995) and listed on May 6, 1997 (62FR24588-24609). Critical habitat was proposed later that year (62FR62741-62751, November 25, 1997) and finally designated on May 5, 1999 (64FR24049-24062) to encompass accessible reaches of all rivers (including estuarine areas and tributaries) between the Mattole River in California and the Elk River in Oregon, inclusive.

The Southern Oregon/Northern California Coast coho salmon ESU occurs between Punta Gorda, Humboldt County, California and Cape Blanco, Curry County, Oregon. Major basins with this salmon ESU are the Rogue, Klamath, Trinity, and Eel river basins, while the Elk River, Oregon, and the Smith and Mad Rivers, and Redwood Creek, California are smaller basins within the range. Hydrologic units and the upstream barriers are Mattole, South Fork Eel, Lower Eel, Middle Fork Eel, Upper Eel (upstream barrier - Scott Dam-Lake Pillsbury), Mad-Redwood, Smith, South Fork Trinity, Trinity (upstream barrier - Lewiston Dam-Lewiston Reservoir), Salmon, Lower Klamath, Scott, Shasta (upstream barrier - Dwinnell Dam-Dwinnell Reservoir), Upper Klamath (upstream barrier - Irongate Dam-Irongate Reservoir), Chetco,

Illinois (upstream barrier - Selmac Dam-Lake Selmac), Lower Rogue, Applegate (upstream barrier - Applegate Dam-Applegate Reservoir), Middle Rogue (upstream barrier - Emigrant Lake Dam-Emigrant Lake), Upper Rogue (upstream barriers - Agate Lake Dam-Agate Lake; Fish Lake Dam-Fish Lake; Willow Lake Dam-Willow Lake; Lost Creek Dam-Lost Creek Reservoir), and Sixes. Related counties are Humboldt, Mendocino, Trinity, Glenn, Lake, Del Norte, Siskiyou in California and Curry, Jackson, Josephine, and Douglas, in Oregon. However, I have excluded Glenn County, California from this analysis because the salmon habitat in this county is not near the agricultural areas where linuron can be used. Klamath county is excluded because it lies beyond an impassable barrier.

Table 40 shows the usage of linuron in the California counties supporting the Southern Oregon/Northern California coastal coho salmon ESU. Table 41 shows the cropping information for Oregon counties where the Southern Oregon/Northern California coastal coho salmon ESU occurs..

Table 40: California Counties where the Southern Oregon/Northern California Coastal Coho Salmon ESU Occurs

County	Site	Acres Treated	Lbs a.i. Applied
Del Norte			None
Humbolt			None
Lake			None
Mendocino			None
Trinity			None

¹ Not a currently registered use.

Table 41. Oregon counties where there is habitat for the Southern Oregon/Northern California coastal coho salmon ESU.

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Curry			None
OR	Douglas			None
OR	Jackson			None
OR	Josephine	Carrot	3	9

There is minimal application of linuron to sites within the Southern Oregon/Northern California Coastal Coho Salmon ESU, and it will have no effect on this ESU.

3. Oregon Coast coho salmon ESU

The Oregon coast coho salmon ESU was first proposed for listing as threatened in 1995 (60FR38011-38030, July 25, 1995), and listed several years later 63FR42587-42591, August 10, 1998). Critical habitat was proposed in 1999 (64FR24998-25007, May 10, 1999) and designated on February 16, 2000 (65FR7764-7787).

This ESU includes coastal populations of coho salmon from Cape Blanco, Curry County, Oregon to the Columbia River. Spawning is spread over many basins, large and small, with higher numbers further south where the coastal lake systems (e.g., the Tenmile, Tahkenitch, and Siltcoos basins) and the Coos and Coquille Rivers have been particularly productive. Critical Habitat includes all accessible reaches in the coastal Hydrologic reaches Necanicum, Nehalem, Wilson-Trask-Nestucca (upstream barrier - McGuire Dam), Siletz-Yaquina, Alsea, Siuslaw, Siltcoos, North Umpqua (upstream barriers - Cooper Creek Dam, Soda Springs Dam), South Umpqua (upstream barrier - Ben Irving Dam, Galesville Dam, Win Walker Reservoir), Umpqua, Coos (upstream barrier - Lower Pony Creek Dam), Coquille, Sixes. Related Oregon counties are Douglas, Lane, Coos, Curry, Benton, Lincoln, Polk, Tillamook, Yamhill, Washington, Columbia, Clatsop. However, the portions of Yamhill, Washington, and Columbia counties that are within the ESU do not include agricultural areas where linuron can be used, and they were eliminated in this analysis.

Table 42: Oregon counties where the Oregon coast coho salmon ESU occurs.

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Benton			None
OR	Clatsop			None
OR	Coos			None
OR	Curry			None
OR	Douglas			None
OR	Lane	Carrot	208	624
OR	Lincoln			None
OR	Polk			None
OR	Tillamook			None

The coastal location and relatively high water flow rates in the Oregon Coast Coho Salmon ESU appears to minimize the very limited linuron application and I expect no effects.

D. Chum Salmon

Chum salmon, *Oncorhynchus keta*, have the widest natural geographic and spawning distribution of any Pacific salmonid, primarily because its range extends farther along the

shores of the Arctic Ocean. Chum salmon have been documented to spawn from Asia around the rim of the North Pacific Ocean to Monterey Bay in central California. Presently, major spawning populations are found only as far south as Tillamook Bay on the northern Oregon coast.

Most chum salmon mature between 3 and 5 years of age, usually 4 years, with younger fish being more predominant in southern parts of their range. Chum salmon usually spawn in coastal areas, typically within 100 km of the ocean where they do not have surmount river blockages and falls. However, in the Skagit River, Washington, they migrate at least 170 km.

During the spawning migration, adult chum salmon enter natal river systems from June to March, depending on characteristics of the population or geographic location. . In Washington, a variety of seasonal runs are recognized, including summer, fall, and winter populations. Fall-run fish predominate, but summer runs are found in Hood Canal, the Strait of Juan de Fuca, and in southern Puget Sound, and two rivers in southern Puget Sound have winter-run fish.

Redds are usually dug in the mainstem or inside channels of rivers. Juveniles outmigrate to seawater almost immediately after emerging from the gravel that covers their redds. This means that survival and growth in juvenile chum salmon depend less on freshwater conditions than on favorable estuarine and marine conditions.

1. Hood Canal Summer-run chum salmon ESU

The Hood Canal summer-run chum salmon ESU was proposed for listing as threatened, and critical habitat was proposed, in 1998 (63FR11774-11795, March 10, 1998). The final listing was published a year later (63FR14508-14517, March 25, 1999), and critical habitat was designated in 2000 (65FR7764-7787).

Critical habitat for the Hood Canal ESU includes Hood Canal, Admiralty Inlet, and the straits of Juan de Fuca, along with all river reaches accessible to listed chum salmon draining into Hood Canal as well as Olympic Peninsula rivers between Hood Canal and Dungeness Bay, Washington. The Hydrologic units are Skokomish (upstream boundary - Cushman Dam), Hood Canal, Puget Sound, Dungeness-Elwha, in the counties of Mason, Clallam, Jefferson, Kitsap, and Island.

Streams specifically mentioned, in addition to Hood Canal, in the proposed critical habitat Notice include Union River, Tahuya River, Big Quilcene River, Big Beef Creek, Anderson Creek, Dewatto River, Snow Creek, Salmon Creek, Jimmycomelately Creek, Duckabush 'stream', Hamma Hamma 'stream', and Dosewallips 'stream'.

Tables 43: Washington counties where the Hood Canal summer-run chum salmon ESU Occurs.

State	County	Site	Acres Treated	lbs a.i. Applied
WA	Clallum			None
WA	Island			None
WA	Jefferson			None
WA	Kitsap	Carrot	1	3

The Hood Canal is a rather well protected body of water in a largely undeveloped portion of Washington State. It is closed to the south and opens to the Straits of Juan de Fuca in the north. To the west, the back ranges of the Olympic Mountains form a protective crest, while to the east the canal is separated by land from Puget Sound and the developed portions of the Puget Sound Basin. As is seen in Table 43, agricultural use of linuron is minimal. The low population density and largely rural nature of the area encompassing the Hood Canal Summer Run Chum Salmon ESU leads me to believe there will be no effects.

2. Columbia River Chum Salmon ESU

The Columbia River chum salmon ESU was proposed for listing as threatened, and critical habitat was proposed, in 1998 (63FR11774-11795, March 10, 1998). The final listing was published a year later (63FR14508-14517, March 25, 1999), and critical habitat was designated in 2000 (65FR7764-7787).

Critical habitat for the Columbia River chum salmon ESU encompasses all accessible reaches and adjacent riparian zones of the Columbia River (including estuarine areas and tributaries) downstream from Bonneville Dam, excluding Oregon tributaries upstream of Milton Creek at river km 144 near the town of St. Helens. These areas are the Hydrologic units of Lower Columbia - Sandy (upstream barrier - Bonneville Dam, Lewis (upstream barrier - Merlin Dam), Lower Columbia - Clatskanie, Lower Cowlitz, Lower Columbia, Lower Willamette in the counties of Clark, Skamania, Cowlitz, Wahkiakum, Pacific, Lewis, Washington and Multnomah, Clatsop, Columbia, and Washington, Oregon. It appears that there are three extant populations in Grays River, Hardy Creek, and Hamilton Creek.

Table 44: Oregon and Washington counties where the Columbia River chum salmon ESU occurs.

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Clatsop			None
OR	Columbia			None
OR	Multnomah			None

OR	Washington	Carrot	1	3
WA	Clark			None
WA	Cowlitz			None
WA	Lewis			None
WA	Pacific			None
WA	Skamania			None
WA	Wahkiakum			None

The Columbia River Chum Salmon ESU is in an area of minimal linuron (3 lb a.i.) application, and there are no effects on the species of interest.

E. Sockeye Salmon

Sockeye salmon, *Oncorhynchus nerka*, are the third most abundant species of Pacific salmon, after pink and chum salmon. Sockeye salmon exhibit a wide variety of life history patterns that reflect varying dependency on the fresh water environment. The vast majority of sockeye salmon typically spawn in inlet or outlet tributaries of lakes or along the shoreline of lakes, where their distribution and abundance is closely related to the location of rivers that provide access to the lakes. Some sockeye, known as kokanee, are non-anadromous and have been observed on the spawning grounds together with their anadromous counterparts. Some sockeye, particularly the more northern populations, spawn in mainstem rivers.

Growth is influenced by competition, food supply, water temperature, thermal stratification, and other factors, with lake residence time usually increasing the farther north a nursery lake is located. In Washington and British Columbia, lake residence is normally 1 or 2 years. Incubation, fry emergence, spawning, and adult lake entry often involve intricate patterns of adult and juvenile migration and orientation not seen in other *Oncorhynchus* species.

Upon emergence from the substrate, lake-type sockeye salmon juveniles move either downstream or upstream to rearing lakes, where the juveniles rear for 1 to 3 years prior to migrating to sea. Smolt migration typically occurs beginning in late April and extending through early July.

Once in the ocean, sockeye salmon feed on copepods, euphausiids, amphipods, crustacean larvae, fish larvae, squid, and pteropods. They will spend from 1 to 4 years in the ocean before returning to freshwater to spawn. Adult sockeye salmon home precisely to their natal stream or lake. River-and sea-type sockeye salmon have higher straying rates within river systems than lake-type sockeye salmon.

1. Ozette Lake Sockeye Salmon ESU

The Ozette Lake sockeye salmon ESU was proposed for listing, along with proposed critical habitat in 1998 (63FR11750-11771, March 10, 1998). It was listed as threatened on March 25, 1999 (64FR14528-14536), and critical habitat was designated on February 16, 2000 (65FR7764-7787). This ESU spawns in Lake Ozette, Clallam County, Washington, as well as in its outlet stream and the tributaries to the lake. It has the smallest distribution of any listed Pacific salmon.

While Lake Ozette, itself, is part of Olympic National Park, its tributaries extend outside park boundaries, much of which is private land. There is limited agriculture in the whole of Clallam County, and most of this is well away from the Ozette watershed.

Table 45: Clallum County where there is habitat for the Ozette Lake sockeye salmon ESU.

State	County	Site	Acres Treated	lbs a.i. Applied
WA	Clallum	Carrot	1	3

The Ozette Lake Sockeye Salmon ESU is located in a remote area of the most northwest county in Washington. There is minimal agriculture and most is located close to the large towns (i.e. Port Angeles). Ozette Lake is protected and located in a largely undeveloped area where tourism is a major industry. There will be no effects from linuron in this ESU.

2. Snake River Sockeye Salmon ESU

The Snake River sockeye salmon was the first salmon ESU in the Pacific Northwest to be listed. It was proposed and listed in 1991 (56FR14055-14066, April 5, 1991 & 56FR58619-58624, November 20, 1991). Critical habitat was proposed in 1992 (57FR57051-57056, December 2, 1992) and designated a year later (58FR68543-68554, December 28, 1993) to include river reaches of the mainstem Columbia River, Snake River, and Salmon River from its confluence with the outlet of Stanley Lake down stream, along with Alturas Lake Creek, Valley Creek, and Stanley, Redfish, Yellow Belly, Pettit, and Alturas lakes (including their inlet and outlet creeks).

Spawning and rearing habitats are considered to be all of the above-named lakes and creeks, even though at the time of the Critical Habitat Notice, spawning only still occurred in Redfish Lake. These habitats are in Custer and Blaine counties in Idaho. However, the habitat area for the salmon is at high elevation, above the agriculture zone, and in protected areas of a National Wilderness area and National Forest. Linuron cannot be used on such a site, and therefore there will be no exposure in the spawning and rearing habitat. There is a probability that this salmon ESU could be exposed to linuron in the lower and larger river reaches during its juvenile or adult migration.

Table 46 shows the limited acreage of crops in Idaho counties where this ESU reproduces. All of this crop production is away from and at a much lower elevation than the spawning and rearing habitat. The critical spawning zones demonstrate no obvious sites for linuron use.

Table 46 shows the acreage of crops where Linuron can be used in Oregon and Washington counties along the migratory corridor for this ESU.

Table 46. Idaho counties where there is spawning and rearing habitat for the Snake River sockeye salmon ESU.

State	County	Site	Acres Treated	lbs a.i. Applied
ID	Blaine			None
ID	Custer			None

Table 47. Oregon and Washington counties that are in the migratory corridors for the Snake River sockeye salmon ESU.

State	County	Site	Acres Treated	lbs a.i. Applied
OR	Clatsop			None
OR	Columbia			None
OR	Gilliam			None
OR	Hood River			None
OR	Morrow	Winter Wheat	1339	670
OR	Multnomah			None
OR	Sherman			None
OR	Umatilla	Asparagus	196	784
OR	Wallowa			None
OR	Wasco			None
WA	Asotin			None
WA	Benton	Asparagus	295	1180
WA	Clark			None
WA	Columbia			None

WA	Franklin	Asparagus	1550	6200
WA	Franklin	Carrot	2752	8256
WA	Garfield			None
WA	Klickitat			None
WA	Walla Walla	Asparagus	255	1020
WA	Pacific			None
WA	Skamania			None
WA	Whitman			None

Although the migratory passages of the Snake River Sockeye Salmon ESU includes many areas of significant agricultural use, the T&E species are more likely to be in larger, downstream rivers and tributaries. The important spawning and rearing areas are at a higher elevation than the agricultural sites, and therefore will not be exposed to the pesticide. It should also be noted that the principal spawning area (Redfish Lake) is located on controlled parklands and not within an area of commercial agriculture. There will be no effects from linuron use to the fish of concern.

4. Specific Conclusions for California and Pacific Northwest Steelhead and Salmon ESUs

The evaluation of linuron by EFED indicated that there were exceedences of the LOC's for linuron. The LOC for endangered species was exceeded for aquatic invertebrates for all sites. The endangered species LOC was exceeded for fresh water fish for wheat, asparagus, and one model of ROW use. This observation suggests some potential for direct effects on the fish species, but indirect effects through loss of the food supply, is more likely. The young salmon and steelhead do not, however, actively feed until movement from the redds is initiated, instead using stored yolk sac material. After active movement begins, the persistence and noted toxicity of linuron to the macroinvertebrate food source may place negative pressure on the young fish, resulting in higher mortality (probably from predation by hatchery fish) and slower growth. Direct, acute, toxicity to fish is slight to moderate.

In addition to being toxic, linuron degrades somewhat slowly and tends to adsorb to the substrate suggesting that any contamination of the water used by endangered salmon and steelhead will remain for some time. Linuron use within the ESU's is moderate in most areas. Particularly in the Pacific Northwest, the major crops (wheat, asparagus, carrots) occupy very large proportions of the land used for agriculture.

The ESUs (7) identified as being potentially affected are situated in areas where major cultivation occurs and this is associated with modest linuron application, but over large areas. These sites are also grouped rather closely, increasing the possibility of higher river

concentrations of pesticide through runoff. Runoff may be significant since label guidelines call for rain or irrigation soon after application for the product to be effective. A final factor in this designation is the presence of spawning and rearing areas for steelhead and chinook salmon. Because the most likely adverse effect from linuron is seen as a reduction in aquatic invertebrates, this indirect effect would be the most noticeable effect.

I again note that the below listed determinations presumed no use of linuron on the wheat crops in the Pacific Northwest. Because of the size of these sites, there is a potential for very large quantities of linuron to be used. This application site, if active, would severely affect several of the ESU's, particularly in the Columbia, Snake, and, potentially, the Willamette Rivers.

Based on these observations, the table below (Table 48) summarizes my conclusions for linuron in California and Northwest Steelhead and Salmon ESU's:

Table 48: Summary of Findings for California and Pacific Northwest Salmon and Steelhead ESUs

Species	ESU	Finding
Chinook Salmon	Upper Columbia	May Affect, Not Likely to Adversely Affect
Chinook Salmon	Snake River spring/summer run	May Affect, Not Likely to Adversely Affect
Chinook Salmon	Snake River fall run	May Affect, Not Likely to Adversely Affect
Chinook Salmon	Upper Willamette	No Effect
Chinook Salmon	Lower Columbia	No Effect
Chinook Salmon	Puget Sound	No Effect
Chinook Salmon	California Coastal	No Effect
Chinook Salmon	Central Valley spring run	No Effect
Chinook Salmon	Sacramento River winter run	No Effect
Coho Salmon	Oregon Coast	No Effect
Coho Salmon	Southern Oregon/Northern California	No Effect
Coho Salmon	Central California	No Effect
Chum Salmon	Hood Canal summer run	No Effect

Chum Salmon	Columbia River	No Effect
Sockeye Salmon	Ozette Lake	No Effect
Sockeye Salmon	Snake River	No Effect
Steelhead	Snake River Basin	No Effect
Steelhead	Upper Columbia River	May Affect, Not Likely to Adversely Affect
Steelhead	Middle Columbia River	May Affect, Not Likely to Adversely Affect
Steelhead	Lower Columbia River	No Effect
Steelhead	Upper Willamette River	No Effect
Steelhead	Northern California	No Effect
Steelhead	Central California Coast	No Effect
Steelhead	South-Central California Coast	May Affect, Not Likely to Adversely Affect
Steelhead	Southern California	May Affect, Not Likely to Adversely Affect
Steelhead	Central Valley California	No Effect

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Attachment 1

Reregistration Eligibility Decision for Linuron

Attachment 2

EFED Linuron RED Chapter

Attachment 3
EPA Quantitative Use Analysis
Linuron

Attachment 4 Sample Labels Linuron

Attachment 5

USGS Distribution of Linuron Use